Supporting Information for Enantioselective Total Synthesis of (–)-Myrifabral A and B Tyler J. Fulton,^a Anthony Y. Chen,^a Michael D. Bartberger,^b and Brian M. Stoltz^{*,a}

 ^aWarren and Katharine Schlinger Laboratory of Chemistry and Chemical Engineering, Division of Chemistry and Chemical Engineering, California Institute of Technology, Pasadena, California 91125, United States stoltz@caltech.edu
^b1200 Pharma LLC, 844 East Green Street, Suite 204, Pasadena, CA, 91101, USA michael.bartberger@1200pharma.com

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Materials and Methods

Unless otherwise stated, reactions were performed in flame-dried glassware under an argon or nitrogen atmosphere using dry, deoxygenated solvents. Solvents were dried by passage through an activated alumina column under argon.¹ Reaction progress was monitored by thin-layer chromatography (TLC) or Agilent 1290 UHPLC-MS. TLC was performed using E. Merck silica gel 60 F254 precoated glass plates (0.25 mm) and visualized by UV fluorescence quenching, panisaldehyde, oxr KMnO₄ staining. Silicycle SiliaFlash® P60 Academic Silica gel (particle size 40–63 µm) was used for flash chromatography. ¹H NMR spectra were recorded on Varian Inova 500 MHz and Oxford 600 MHz spectrometers and are reported relative to residual CHCl₃ ($\delta = 7.26$ ppm) or TMS ($\delta = 0.00$ ppm). ¹³C NMR spectra were recorded on a Bruker 400 MHz spectrometer (100 MHz) and are reported relative to CHCl₃ (δ = 77.16 ppm), C₆D₆ (δ = 128.06 ppm). Data for ¹H NMR are reported as follows: chemical shift (δ ppm) (multiplicity, coupling constant (Hz), integration). Multiplicities are reported as follows: s = singlet, d = doublet, t = triplet, q = quartet, p = pentet, sept = septuplet, m = multiplet, br s = broad singlet, br d = broad doublet. Data for ${}^{13}C$ NMR are reported in terms of chemical shifts (δ ppm). IR spectra were obtained by use of a Perkin Elmer Spectrum BXII spectrometer or Nicolet 6700 FTIR spectrometer using thin films deposited on NaCl plates and reported in frequency of absorption (cm⁻¹). Optical rotations were measured with a Jasco P-2000 polarimeter operating on the sodium D-line (589 nm), using a 100 mm pathlength cell. High resolution mass spectra (HRMS) were obtained from Agilent 6200 Series TOF with an Agilent G1978A Multimode source in electrospray ionization (ESI+), atmospheric pressure chemical ionization (APCI+), or mixed ionization mode (MM: ESI-APCI+). Reagents were purchased from commercial sources and used as received unless otherwise stated.

List of Abbreviations:

TLC – thin-layer chromatography VCD – vibrational circular dichroism PHOX – phosphinooxazoline



α-aminomethyl β-keto ester (11)

Compound 11 was prepared as previously described by Stoltz et al.² To a stirred solution of β keto ester 9 (20.0 g, 120.3 mmol, 1.0 equiv) in CH₂Cl₂ (600 mL) was added sulforylmethyl carbamate 10 (39.2 g, 144.4 mmol, 1.2 equiv) in one portion at ambient temperature. After stirring for 5 min, Cs₂CO₃ (98.0 g, 300 mmol, 2.5 equiv) was added in one portion. The resulting white suspension was stirred vigorously at 20 °C. After 16 h, full consumption of starting material was determined by TLC analysis. Saturated aqueous ammonium chloride (300 mL) was added slowly, and the biphasic mixture was stirred at ambient temperature for 40 min and extracted with CH₂Cl₂ (3 x 100 mL). The combined organic layers were dried over Na₂SO₄, filtered, and concentrated to a viscous, colorless oil. Flash column chromatography (SiO₂, 10% EtOAc in hexanes) afforded α aminomethyl β-keto ester 11 as an amorphous white solid (35.61 g, 114.36 mmol, 95% yield); ¹H NMR (500 MHz, CDCl₃) δ 5.91 (ddt, J = 17.2, 10.4, 5.9, 1H), 5.33 (dq, J = 17.2, 1.4, 1H), 5.25 (dd, J = 10.4, 1.4 Hz, 1H), 5.17 (t, J = 5.7 Hz, 1H), 4.63 (d, J = 5.8 Hz, 1H), 3.55 (dd, J = 13.9)7.7 Hz, 1H), 3.41 (dd, J = 13.9, 5.7 Hz, 1H), 2.63–2.34 (m, 3H), 2.07–1.94 (m, 1H), 1.87–1.75 (m, 1H), 1.74 – 1.53 (m, 4H), 1.41 (s, 9 H);¹³C NMR (100 MHz, CDCl₃) δ 208.9, 170.9, 155.9, 131.6, 119.2, 79.4, 66.3, 62.4, 44.3, 40.8, 33.8, 28.4, 27.2, 22.1; IR (Neat Film, NaCl) 3460, 2936, 2869, 1712, 1501, 1451, 1391, 1366, 1315, 1225, 1202, 1170, 1140, 1099, 931, 856 cm⁻¹; HRMS (MM:ESI-APCI+) m/z calc'd for C₁₆H₂₆NO₅ [M+H]⁺: 312.1805, found 312.1805.



Glutarimide 8

A flame-dried 2 L round bottom flask equipped with a Dean–Stark trap, a reflux condenser and a stirring bar was charged with **11** (15.00 g, 48.17 mmol, 1.0 equiv), glutaric acid (12.73 g, 96.35 mmol, 2.0 equiv), 4-CF₃PhB(OH)₂ (915.3 mg, 4.82 mmol, 10 mol %), and xylenes (960 mL). The resulting suspension was heated to reflux in a heating mantle with vigorous stirring. After 48 h, the light orange reaction solution was cooled to 20 °C and concentrated to provide a crude orange oil. The crude oil was purified by column chromatography ($2 \rightarrow 5\%$ EtOAc in CH₂Cl₂) to afford an off white semisolid which was further purified by column chromatography (SiO₂, $20\rightarrow 40\%$ EtOAc in hexanes) to provide **8** as an amorphous white solid (12.62 g, 41.05 mmol, 85% yield); ¹H NMR (500 MHz, CDCl₃) δ 5.94 (dddd, J = 17.3, 10.4, 6.1, 5.6 Hz, 1H), 5.33 (dd, J = 17.3, 1.5 Hz, 1H), 5.25 (dd, J = 10.4, 1.5 Hz, 1H), 4.66 (ddt, J = 13.1, 6.1, 1.2 Hz, 1H), 4.53 (ddt, J = 13.1, 5.6, 1.2 Hz, 1H), 4.32 (ABq, $\Delta \delta_{AB} = 0.05$, $J_{AB} = 14.1$ Hz, 2H), 2.64 (t, J = 6.5 Hz, 4H), 2.48–2.36 (m, 2H), 2.30–2.24 (m, 1H), 2.03–1.97 (m, 1H), 1.94 (p, J = 6.6 Hz, 2H), 1.77–1.70 (m, 1H), 1.67–1.49 (m, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 206.0, 172.8, 169.8, 131.5, 118.6, 66.4, 59.6, 41.9, 40.9, 34.0, 32.8, 27.1, 22.2, 16.7; IR (Neat Film, NaCl) 2942, 17111, 1681, 1426, 1379, 1334, 1265, 1204, 1122, 1098, 1057, 1020 cm⁻¹; HRMS (MM:ESI-APCI+) *m/z* calc'd for C₁₆H₂₂NO₅ [M+H]⁺: 308.1492, found 308.1504.



Ketone 7

An oven-dried 1 L round bottom flask was charged with Pd₂(dba)₃ (819.6 mg, 0.895 mmol, 2.75 mol %), (*S*)-(CF₃)₃-*t*-BuPHOX (1.1548 g, 1.95 mmol, 6.0 mol %), and a magnetic stirring bar in a N₂-filled glovebox. The flask was then charged with toluene (650 mL) and stirred at 24 °C for 40 min, generating a dark orange/red solution. The preformed catalyst solution was then cannulated into a solution of **8** (10.0 g, 32.54 mmol, 1.0 equiv) dissolved in toluene (325 mL) in a 2 L flame-dried round bottom flask. The resulting dark green solution was stirred at 24 °C. Full consumption of the starting material was achieved after 7 h, as determined by TLC analysis (25% EtOAc in hexanes). The crude reaction mixture was concentrated and directly purified by column chromatography (SiO₂, 10 \rightarrow 40% EtOAc in hexanes) to yield glutarimide 7 as an off white semisolid (8.06 g, 30.6 mmol, 94% yield); 88% ee, $[\alpha]_D^{25}$ +32.9 (*c* 1.0, CHCl₃); ¹H NMR (500 MHz, C₆D₆) δ 5.67 (dddd, *J* = 16.7, 10.3, 8.4, 5.9 Hz, 1H), 5.04–4.94 (m, 2H), 4.12 (ABq, $\Delta \delta_{AB} = 0.22$,

 $J_{AB} = 13.7$ Hz, 2H), 2.90 (ddd, J = 15.6, 12.4, 6.2 Hz, 1H), 2.63 (t, J = 6.4 Hz, 4H), 2.38 (ddt, J = 14.2, 5.8, 1.3 Hz, 1H), 2.32 (dt, J = 15.8, 4.4 Hz, 1H), 2.01–1.87 (m, 4H), 1.83–1.68 (m, 4H), 1.64–1.53 (m, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 213.2, 173.2, 134.7, 118.2, 51.8, 44.2, 39.6, 38.7, 35.3, 33.1, 26.1, 21.2, 16.8; IR (Neat Film, NaCl) 3072, 2937, 2864, 1727, 1701, 1679, 1638, 1430, 1380, 1340, 1319, 1243, 1173, 1120, 1056, 1008, 914, 868, 803 cm⁻¹; HRMS (MM:ESI-APCI+) *m/z* calc'd for C₁₅H₂₂NO₃ [M+H]⁺: 264.1594, found 264.1591; SFC Conditions: 35% IPA, 3.5 mL/min, Chiralpak AD-H column, $\lambda = 210$ nm, t_R (min): major = 1.28, minor = 1.68.



Determination of Absolute Configuration of 7 and ent-7.

Method 1 – Vibrational Circular Dichroism (VCD)

Experimental Protocol. Solutions of compounds 7 and *ent*-7 (69 mg/mL) were each prepared in CDCl₃ and loaded into a front-loading SL-4 cell (International Crystal Laboratories) possessing BaF₂ windows and 100 μ m path length. Infrared (IR) and VCD spectra were individually acquired on a BioTools ChiralIR-2X VCD spectrometer as sets of 24 one-hour blocks (24 blocks, 3120 scans per block) at 4 cm⁻¹ resolution in dual PEM mode. A 15-minute acquisition of neat (–)- α -pinene control (separate 75 μ m BaF₂ cell) yielded a VCD spectrum in agreement with literature spectra and those previously acquired on the same instrument. IR and VCD spectra were background-corrected using a 30-minute block IR acquisition of the empty instrument chamber under gentle N₂ purge, and were solvent corrected using a 12-hour (12 blocks, 3120 scans per

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block) IR/VCD acquisition of CDCl₃ in the same 100 μ m BaF₂ cell as used for 7 and *ent*-7. The reported spectra represent the result of block averaging.

Computational Protocol. The arbitrarily chosen (R) enantiomer of compound 7 was subjected to an initial exhaustive stochastic molecular mechanics-based conformational search (MMFF94 force field, 0.06 Å geometric RMSD cutoff, and 30 kcal/mol energy window) as implemented in MOE 2019.0102 (Chemical Computing Group, Montreal, CA). All conformers retained the (R)configuration and were subjected to geometry optimization, harmonic frequency calculation, and VCD rotational strength evaluation using density functional theory. Initial quantum mechanical calculations utilized the B3LYP functional, small 6-31G* basis, and IEFPCM model (chloroform solvent) as an initial filter. This was followed by subsequent treatment using the B3PW91 functional, cc-pVTZ basis, and implicit IEFPCM chloroform solvation model on all IEFPCM-B3LYP/6-31G* conformers below 5 kcal/mol, reusing the exact Hessian of the latter to facilitate optimization at the higher level of theory. All calculations were performed with the Gaussian 16 program system (Rev. C.01; Frisch et al., Gaussian, Inc., Wallingford, CT). Resultant IEFPCM-B3PW91/cc-pVTZ harmonic frequencies were scaled by 0.98. All structurally unique conformers possessing all positive Hessian eigenvalues were Boltzmann weighted by relative free energy at 298.15 K. The predicted IR and VCD frequencies and intensities of the retained conformers were convolved using Lorentzian line shapes ($\gamma = 4 \text{ cm}^{-1}$) and summed using the respective Boltzmann weights to yield the final predicted IR and VCD spectra. The predicted VCD spectrum of the (S) enantiomer was generated by inversion of sign. From the outstanding agreement between the theoretical and measured IR and VCD spectra across the entire useful range of the spectrum (900– 1500 cm⁻¹; regions A–J below) along with support of this assignment using the directly predicted versus measured optical rotations (see Method 2) the absolute configurations of species 7 and ent-7 were established as (S) and (R), respectively.



Experimental (left) and computed (right) IR and VCD spectra of 7 and ent-7.

Method 2 – Optical Rotation (OR)

Computational Protocol. The ensemble of unique IEFPCM-B3PW91/cc-pVTZ conformers of (R)-7 generated in Method 1 above were subjected to optical rotation calculation at 589.0 nm using the B3LYP hybrid density functional, the large and diffuse 6-311++G(2df,2pd) basis set, and the IEFPCM implicit chloroform solvent model. The computed IEFPCM-B3LYP/6-311++G(2df,2pd) optical rotations (weighted by IEFPCM-B3PW91/cc-pVTZ free energies at 298.15 K) along with those resulting from alternatively weighting by either the IEFPCM-B3PW91/cc-pVTZ IEFPCM-B3LYP/6-311++G(2df,2pd)//IEFPCMtotal energies or B3PW91/cc-pVTZ total energies are reported in (a)-(b) below. From comparison of the theoretically calculated and measured optical rotations (for which reasonably good agreement in magnitude was found to exist) the respective VCD-based AC assignments of (S) and (R) for 7 and ent-7 were further supported by those from the separate OR-based methodology. The individual

relative energies, free energies, and optical rotation signatures of each conformer of (R)-7 are provided in the accompanying Microsoft Excel file.

Predicted optical rotation, weighted by IEFPCM-B3PW91/cc-pVTZ free energies: -47.5° Predicted optical rotations, weighted by IEFPCM-B3PW91/cc-pVTZ total energies: -45.5° Predicted optical rotations, weighted by IEFPCM-B3LYP/6-311++G(2df,2pd)//IEFPCM-B3PW91/cc-pVTZ total energies: -45.8°



Predicted optical rotation, weighted by IEFPCM-B3PW91/cc-pVTZ free energies: +47.5° Predicted optical rotations, weighted by IEFPCM-B3PW91/cc-pVTZ total energies: +45.5° Predicted optical rotations, weighted by IEFPCM-B3LYP/6-311++G(2df,2pd)//IEFPCM-B3PW91/cc-pVTZ total energies: +45.8°

Measured optical rotation: (CHCl₃ solvent, 25 °C, c = 10.0 mg/mL, 10 cm pathlength, 88% ee) 7: + 32.9 *ent*-7: -28.9



Ethyl vinyl ether 14

A flame-dried 250 mL round bottom flask was charged with 7 (3.00 g, 11.4 mmol, 1.0 equiv), EtOH (114 mL), p-TsOH•H₂O (43.4 mg, 0.228 mmol, 0.02 equiv), and CH(OEt)₃ (38.0

mL, 228 mmol, 20.0 equiv). The resulting clear, colorless solution was heated in a 40 °C heating block for 16 h, after which time complete conversion was observed by TLC analysis (40% EtOAc in hexanes). The reaction mixture was concentrated under reduced pressure and the resulting colorless oil was dissolved in EtOAc (50 mL) and poured into a separatory funnel containing saturated aqueous NaHCO₃ (50 mL). The layers were separated and the aqueous layer was extracted with EtOAc (2 x 100 mL). The combined organics were washed with brine (25 mL), dried over Na₂SO₄, filtered, and concentrated. Flash column chromatography (SiO₂, 30% EtOAc in hexanes) provided **14** as a colorless oil (3.0326 g, 10.41 mmol, 91% yield); $[\alpha]_D^{25}$ -72.6 (c 1.0, CHCl₃); ¹H NMR (500 MHz, C₆D₆) δ 5.90 (dddd, J = 17.3, 10.0, 9.2, 5.3 Hz, 1H), 5.18–5.01 (m, 2H), 4.66–4.54 (m, 1H), 4.35 (ABq, $\Delta \delta_{AB} = 0.23$, $J_{AB} = 13.1$ Hz, 2H), 3.55–3.31 (m, 2H), 2.95 (ddt, J = 13.4, 5.1, 1.3 Hz, 1H), 2.14–1.81 (m, 8H), 1.77–1.64 (m, 1H), 1.64–1.50 (m, 2H), 1.10 $(t, J = 7.0 \text{ Hz}, 3\text{H}), 1.06 - 0.91 \text{ (m, 2H)}; {}^{13}\text{C} \text{ NMR} (100 \text{ MHz}, C_6D_6) \delta 172.2, 156.5, 136.4, 117.1,$ 96.6, 61.8, 45.1, 42.7, 41.4, 33.2, 31.8, 24.5, 19.5, 16.8, 14.8; IR (Neat Film, NaCl) 3393, 3071, 2974, 2935, 2876, 2839, 1730, 1682, 1430, 1359, 1341, 1275, 1240, 1220, 1176, 1158, 1138, 1113, 1057, 1046, 1002, 930, 912, 879, 846, 816, 787, 745, 698 cm⁻¹; HRMS (MM:ESI-APCI+) m/z calc'd for C₁₇H₂₆NO₃ [M+H]⁺: 292.1907, found 292.1910.



Tricyclic lactam 6

A flame-dried 250 mL round bottom flask was charged with **14** (2.50 g, 8.58 mmol, 1.0 equiv) and CH₂Cl₂ (86 mL). The resulting clear, colorless solution was cooled in a -78 °C bath. After 15 min, LiEt₃BH in (9.44 mL, 1.0 M in THF, 9.44 mmol, 1.1 equiv) was added dropwise over 5 min. After 30 min, an additional portion of LiEt₃BH (360 µL, 0.360 mmol, 0.042 equiv) was added. After 10 min, an additional portion of LiEt₃BH (300 µL, 0.300 mmol, 0.035 equiv) was added. After stirring for 10 min, the reaction was complete by TLC analysis (40% EtOAc in hexanes). EtOAc (210 µL, 2.15 mmol, 0.25 equiv) was added and the reaction was allowed to stir at -78 °C. After 1 h, BF₃•OEt₂ (2.11 mL, 17.2 mmol, 2.0 equiv) was added dropwise over 5 min. After 15 min, the reaction was complete by LC/MS analysis. The reaction mixture was quenched with H₂O (60 mL), warmed to 20 °C, and transferred to a separatory funnel with CH₂Cl₂ (10 mL).

The layers were separated and the aqueous was extracted with CH₂Cl₂($3 \times 25 \text{ mL}$). The combined organics were dried over Na₂SO₄, filtered, and concentrated to afford a pale yellow oil. Purification by column chromatography (SiO₂, 25% acetone in hexanes) yielded **6** as a white semisolid (1.8841 g, 7.62 mmol, 89% yield); $[\alpha]_D^{25}$ –22.6 (*c* 1.0, CHCl₃); ¹H NMR (400 MHz, C₆D₆) δ 5.91 (ddt, *J* = 17.4, 10.2, 7.5 Hz, 1H), 5.06–4.93 (m, 3H), 2.97 (dtd, *J* = 10.6, 5.5, 2.8 Hz, 1H), 2.49 (dd, *J* = 13.7, 2.4 Hz, 1H), 2.29 (ddt, *J* = 17.1, 4.8, 2.6 Hz, 1H), 2.18 (dq, *J* = 7.4, 1.3 Hz, 2H), 1.93 (dq, *J* = 4.9, 2.7 Hz, 1H), 1.91–1.72 (m, 4H), 1.48–1.30 (m, 2H), 1.25 – 1.15 (m, 1H), 1.15–1.05 (m, 1H), 1.04 – 0.95 (m, 2H), 0.94–0.85 (m, 1H); ¹³C NMR (100 MHz, C₆D₆) δ 213.8, 168.5, 134.1, 118.3, 59.9, 52.8, 51.6, 49.1, 39.6, 39.2, 33.3, 28.9, 26.7, 20.5, 19.7; IR (Neat Film, NaCl) 3074, 2937, 2870, 1716, 1644, 1460, 1440, 1416, 1346, 1264, 1235, 1166, 1120, 1064, 994, 958, 916 cm⁻¹; HRMS (MM:ESI-APCI+) *m/z* calc'd for C₁₅H₂₁NO₂ [M+H]⁺: 248.1645, found 248.1653.



Amino alcohol 16

A two-necked 250 mL round bottom flask equipped with a reflux condenser, septum, and stir bar was charged with **6** (905 mg, 3.66 mmol, 1.0 equiv) and THF (73 mL). A flame-dried 25 mL conical flask under N₂ was charged with L-Selectride (5.90 mL, 1.0 M in THF, 5.90 mmol, 1.6 equiv). Both flasks were cooled to -78 °C for 30 min, after which time the L-Selectride solution was slowly transferred to the flask containing **6** via syringe over 20 min via positive pressure cannulation, resulting in the formation of an opaque white reaction mixture. After 30 min, the septum was exchanged for an oven-dried glass stopper and LiAlH₄ (555.6 mg, 14.6 mmol, 4.0 equiv) was added in a single portion to the reaction mixture. The reaction was then removed from the cooling bath and allowed to reach 20 °C, after which time the flask was immersed in a 75 °C oil bath. After refluxing for 9 h, complete conversion to **16** was observed by LC/MS and TLC analysis. The reaction mixture was diluted with Et₂O (50 mL) and cooled in an ice/water bath. After 10 min, the reaction was slowly quenched with dropwise addition of H₂O (800 µL) over 10 min, followed by the addition of aqueous NaOH (4.0 mL of a 2.5 M solution), and H₂O₂ (4.0 mL of a 30% solution). The resulting gray suspension was then stirred vigorously for 30 min before being filtered through a pad of celite (5 x 5 cm), washing with EtOAc (3 x 75 mL). The filtrate

was then transferred to a separatory funnel and washed with H₂O (60 mL) and brine (60 mL). The combined aqueous layers were extracted with EtOAc (3 x 80 mL). The combined organics were then dried over Na₂SO₄, filtered, and concentrated to a pale yellow oil. Purification by column chromatography (SiO₂, 25% EtOAc in hexanes with 1% Et₃N) afforded **16** as a colorless, viscous oil which slowly turns red with exposure to air (833.1 mg, 3.54 mmol, 97% yield); $[\alpha]_D^{25}$ –13.4 (*c* 1.0, CHCl₃); ¹H NMR (500 MHz, C₆D₆) δ 5.83 (dddd, *J* = 15.1, 11.2, 8.3, 7.6 Hz, 1H), 5.04 (app t, *J* = 1.2 Hz, 1H), 5.02–4.99 (m, 1H), 3.25 (m, 1H), 2.78–2.60 (m, 2H), 2.55 (d, *J* = 11.3 Hz, 1H), 2.08–1.74 (m, 6H), 1.68–1.37 (m, 9H), 1.32–1.03 (m, 3H); ¹³C NMR (100 MHz, C₆D₆) δ 134.8, 116.8, 75.4, 65.5, 65.4, 56.4, 43.3, 41.8, 37.8, 30.5, 30.0, 26.1, 24.7, 20.9, 20.0; IR (Neat Film, NaCl) 3404 (br), 3072, 2931, 2856, 2797, 2759, 1638, 1463, 1442, 1375, 1336, 1271, 1223, 1198, 1182, 1124, 1106, 1044, 995, 958, 947, 912, 844, 813, 768, 714, 678, 635 cm⁻¹; HRMS (MM:ESI-APCI+) *m/z* calc'd for C₁₅H₂₆NO [M+H]⁺: 236.2009, found 236.2012.



(–)- α , β -myrifabral A (4)

In a nitrogen-filled glovebox, an oven-dried 25 mL round bottom flask was charged with amino alcohol **16** (100.0 mg, 0.424 mmol, 1.0 equiv), pinacol boronate **17** (356.2 mg, 2.12 mmol, 5.0 equiv), and a Teflon-coated stir bar. The flask was sealed with a septum, removed from the glovebox, and placed under an atmosphere of nitrogen. To the flask was added THF (2.2 mL) to provide a clear, colorless solution. Hoveyda–Grubbs II catalyst (26.6 mg, 0.0424 mmol, 10 mol %) was then added rapidly in a single portion, and the flask was subjected to vacuum until the green solution began to bubble. The dark green reaction was allowed to stir for 10 min under a static vacuum, at which point the flask was backfilled with nitrogen and an aliquot was analyzed by LC/MS, indicating full conversion of amino alcohol **16** to the putative cross metathesis product. *Note: Typically, the reaction solution rapidly turns dark brown/black when the flask is backfilled with nitrogen*. The metathesis catalyst was quenched with the addition of ethyl vinyl ether (40 μ L) at 20 °C. After stirring for 30 min at 20 °C, deionized water (2.2 mL) and NaBO₃•4H₂O (1.30 g, 8.48 mmol, 20.0 equiv) were added and the resulting black, biphasic suspension was stirred rapidly

at 20 °C. After 4 h, full conversion of the intermediate cross metathesis product was observed by LC/MS analysis. The reaction mixture was poured into a separatory funnel with EtOAc (5 mL) and H₂O (10 mL). The layers were separated and the aqueous layer was extracted 3 x 5 mL EtOAc. The combined organics were dried over Na₂SO₄, filtered, and concentrated to yield a dark brown oil. Purification by column chromatography (SiO₂, $0 \rightarrow 50\%$ EtOAc in hexanes with 2% Et₃N) yielded (-)-myrifabral A (4) as a 1.4:1 mixture of β : α -OH epimers as a viscous yellow oil (90.1 mg, 0.358 mmol, 85% yield). Note: racemic samples of this compound are isolated as a colorless solid, in accordance with previous reports.^{3,4} Due to the complicated overlap of β and α -OH epimers of (-)-4, the ¹H NMR spectral data are reported with raw integration values: $\left[\alpha\right]_{D}^{25}$ -41.4 (c 1.0, CHCl₃); ¹H NMR (600 MHz, Pyridine- d_5) δ 8.36 (s, 1.00), 7.70 (s, 0.72), 5.68 (d, J = 3.7Hz, 0.82), 5.10 (d, J = 9.6 Hz, 1.19), 4.19 (d, J = 3.4 Hz, 0.89), 3.21 (d, J = 3.6 Hz, 1.17), 2.80– 2.64 (m, 4.30), 2.54 (t, J = 10.7 Hz, 2.16), 2.30–2.19 (m, 2.22), 2.10 – 1.88 (m, 7.28), 1.88 – 1.80 (m, 2.19), 1.78-1.72 (d, J = 11.9 Hz, 3.43), 1.68-1.59 (m, 8.86), 1.57-1.40 (m, 8.86), 1.27-1.05(m, 10.40); ¹³C NMR (100 MHz, Pyridine-*d*₅) δ 98.3, 92.3, 80.7, 72.5, 69.6, 69.1, 66.8, 66.4, 57.2, 57.2, 40.4, 40.2, 34.4, 33.1, 32.6, 30.7, 30.7, 30.6, 29.6, 29.6, 28.7, 27.5, 26.9, 26.9, 25.5, 25.5, 21.4, 21.3, 21.3, 21.2; IR (Neat Film, NaCl) 3381 (br), 3054, 2933, 2851, 2796, 2756, 2728, 2253, 1714, 1562, 1549, 1540, 1462, 1456, 1444, 1396, 1374, 1336, 1298, 1277, 1243, 1209, 1189, 1124, 1106, 1077, 1057, 1009, 998, 961, 949, 928, 917, 902, 886, 870, 856, 840, 826, 762, 739, 704, 673 cm⁻¹, HRMS (MM:ESI-APCI+) *m/z* calc'd for C₁₅H₂₆NO₂ [M+H]⁺: 252.1958, found 252.1960.





 $(-)-\alpha,\beta$ -myrifabral B (5)

The following procedure was adapted from Song's total synthesis of (±)-myrifabral B.⁴ To a one dram vial with a stir bar was added (–)-4 (50.0 mg, 0.199 mmol, 1.0 equiv), THF (220 μ L), 2 N aqueous HCl (250 μ L), and *N*-ethyl-*N*-(methoxymethyl)ethanamine (117.2 mg, 0.995 mmol, 5.0 equiv) to provide a colorless, biphasic reaction mixture. The vial was sealed with a Teflonlined cap and heated in a vial well at 80 °C with rapid stirring. After 4 h, the reaction was complete by LC/MS analysis. The vial was cooled to 20 °C, then the biphasic reaction mixture was poured into a separatory funnel with EtOAc (5 mL), and saturated aqueous NaHCO₃ (10 mL). The layers were separated and the aqueous layer was extracted with EtOAc (3 x 5 mL). The combined organics were dried over Na₂SO₄, filtered, and concentrated to a brown/yellow oil. Purification by column chromatography (SiO₂, 20 \rightarrow 60% EtOAc in hexanes with 2% Et₃N) yielded (–)myrifabral B (**5**) as a 1.6:1 mixture of β : α -OH epimers as a viscous yellow oil (29.5 mg, 0.0877 mmol, 44% yield). Note: racemic samples of this compound are isolated as a colorless solid, in accordance with previous reports.^{3,4} Due to the complicated overlap of β and α-OH epimers of (–)-5, the ¹H NMR spectral data are reported with raw integration values; $[α]_D^{25}$ –37.5 (*c* 1.0, CHCl₃); ¹H NMR (600 MHz, Pyridine-*d*₅) δ 5.72 (d, *J* = 3.0 Hz, 1.00), 4.82 (d, *J* = 8.2 Hz, 1.67), 4.22 (d, *J* = 3.4 Hz, 1.01), 4.22 (d, *J* = 3.4 Hz, 1.40), 2.83–2.74 (m, 2.40), 2.73–2.68 (m, 3.86), 2.66 (dd, *J* = 12.7, 7.9 Hz, 1.75), 2.60 (dd, *J* = 10.9, 7.2 Hz, 2.92), 2.57–2.47 (m, 7.18), 2.45–2.33 (m, 6.41), 2.28 (tdd, *J* = 13.1, 6.6, 2.4 Hz, 1.64), 2.21 (dd, *J* = 12.8, 6.1 Hz, 1.61), 2.15–2.01 (m, 4.23), 2.01–1.93 (m, 4.26), 1.83–1.78 (m, 3.00), 1.73–1.60 (m, 9.07), 1.58–1.43 (m, 11.78), 1.39 (dd, *J* = 13.3, 4.4 Hz, 1.56), 1.34–1.28 (m, 3.61), 1.25 (dt, *J* = 13.0, 3.1 Hz, 2.86), 1.21–1.09 (m, 3.00), 1.02 (t, *J* = 7.1 Hz, 5.89), 0.99 (t, *J* = 7.1 Hz, 8.22), 0.85 (t, *J* = 12.9 Hz, 1.39); ¹³C NMR (100 MHz, Pyridine-*d*₅) δ 103.1, 94.2, 80.4, 72.6, 69.8, 69.0, 66.9, 66.4, 57.2, 57.2, 57.1, 56.8, 48.5, 48.0, 40.1, 40.0, 39.1, 37.3, 35.2, 35.2, 33.8, 33.8, 30.7, 30.7, 30.5, 29.8, 26.9, 26.8, 25.5, 25.4, 21.6, 21.4, 21.3, 21.3, 12.9, 12.4; IR (Neat Film, NaCl) 3076 (br), 2966, 2933, 2851, 2801, 2757, 2728,

2251, 1722, 1692, 1557, 1462, 1444, 1375, 1346, 1297, 1276, 1267, 1243, 1205, 1196, 1178, 1144, 1124, 1104, 1089, 1076, 1059, 1037, 967, 918, 904, 885, 813, 860, 833, 753, 715, 666, 616 cm⁻¹, HRMS (FAB+) *m/z* calc'd for C₂₀H₃₇N₂O₂ [M+H]⁺: 337.2855, found 337.2857.





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¹³C NMR (100 MHz, CDCl₃) of compound **11**.





¹³C NMR (100 MHz, CDCl₃) of compound **8**.





¹³C NMR (100 MHz, CDCl₃) of compound **7**.

S23





 13 C NMR (100 MHz, C₆D₆) of compound **14**.

S25

92'T













S29





¹³C NMR (100 MHz, pyridine- d_5) of compound (-)-4.





¹³C NMR (100 MHz, pyridine- d_5) of compound (-)-5.