

Supporting Information

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Atroposelective Synthesis of PINAP via Dynamic Kinetic Asymmetric Transformation

Seo-Jung Han, Vikram Bhat, Brian M. Stoltz,* and Scott C. Virgil*

Supporting Information for

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Asymmetric Transformation**

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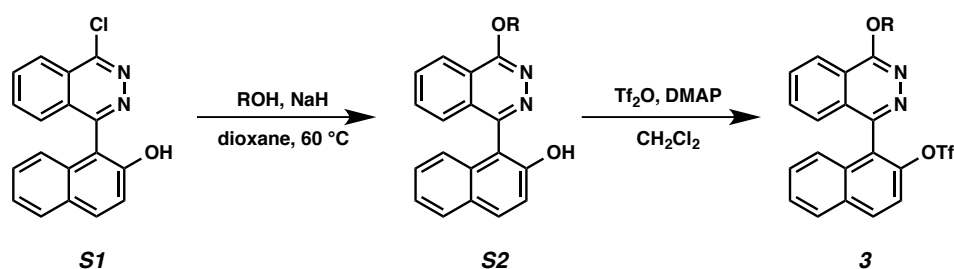
Materials and Methods.

Unless otherwise stated, reactions were performed in flame-dried glassware under an argon or nitrogen atmosphere using dry, deoxygenated solvents. Reaction progress was monitored by thin-layer chromatography (TLC). THF, Et₂O, CH₂Cl₂, toluene, benzene, CH₃CN, and dioxane were dried by passage through an activated alumina column under argon. Purified water was obtained using a Barnstead NANOpure Infinity UV/UF system. Brine solutions are saturated aqueous solutions of sodium chloride. Commercially available reagents were purchased from Sigma-Aldrich, Acros Organics, Strem, or Alfa Aesar and used as received unless otherwise stated. Reaction temperatures were controlled by an IKA Mag temperature modulator unless otherwise indicated. Glove box manipulations were performed under a N₂ atmosphere. TLC was performed using E. Merck silica gel 60 F254 precoated glass plates (0.25 mm) and visualized by UV fluorescence quenching, *p*-anisaldehyde, or PMA (phosphomolybdic acid) staining. Silicycle SiliaFlash P60 Academic Silica gel (particle size 0.040-0.064 mm) was used for flash column chromatography. ¹H NMR spectra were recorded on a Varian Inova 500 MHz spectrometer and are reported relative to residual CHCl₃ (δ 7.26 ppm). ¹³C NMR spectra are recorded on a Varian Inova 500 MHz spectrometer (125MHz) and are reported relative to CHCl₃ (δ 77.16 ppm). Data for ¹H NMR are reported as follows: s = singlet, d = doublet, t = triplet, q = quartet, p = pentet, sept = septuplet, m = multiplet, br s = broad singlet, br d = broad doublet, app = apparent. IR spectra were obtained using a Perkin Elmer Paragon 1000 spectrometer using thin films deposited on NaCl plates and reported in frequency of absorption (cm⁻¹). Optical rotations were measured with a Jasco P-2000 polarimeter operating on the sodium D-line (589 nm), using a 100 mm path-length cell and are reported as: [α]_D^T (concentration in g/100 mL, solvent). Analytical HPLC was performed with an Agilent 1100 Series HPLC utilizing a Chiralpak (AD-H or AS) or Chiralcel (OD-H, OJ-H, or OB-H) columns (4.6 mm x 25 cm) obtained from Daicel Chemical Industries, Ltd. Analytical SFC was performed with a Mettler SFC supercritical CO₂ analytical chromatography system utilizing Chiralpak (AD-H, AS-H or IC) or Chiralcel (OD-H, OJ-H, or OB-H) columns (4.5 mm x 25 cm) obtained from Daicel Chemical Industries, Ltd. High resolution mass spectra (HRMS) were obtained from the Caltech Mass Spectral Facility using JEOL JMS-600H High Resolution Mass Spectrometer in fast atom bombardment (FAB+) or electron

ionization (EI+) mode, or Agilent 6200 Series TOF with an Agilent G1978A Multimode source in electrospray ionization (ESI+), atmospheric pressure chemical ionization (APCI+), or mixed ionization mode (MM: ESI-APCI+).

Triflate substrates **3i–3k** were prepared by known methods.¹ All the data from PINAP **4i–4k** were confirmed by reported data.¹

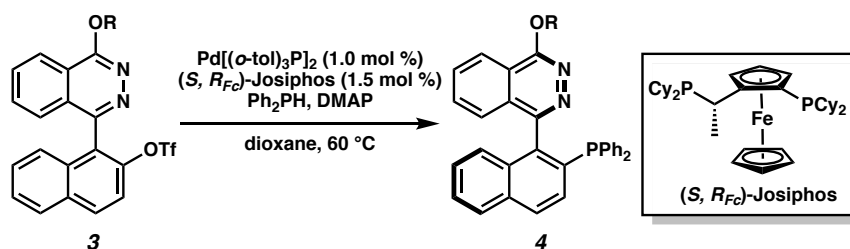
General Procedure for Synthesis of Triflate Substrates



To a solution of NaH (4.00 mmol, 2.00 equiv) in dioxane (8.00 mL) was added alcohol (2.10 mmol, 1.05 equiv) in dioxane (2.00 mL) dropwise. The reaction mixture was stirred for 15 min at 23 °C and then **S1** (2.00 mmol, 1.00 equiv) was added portionwise. The solution was stirred for 18 h at 60 °C then diluted with EtOAc (10.0 mL) and water (20.0 mL). The aqueous phase was extracted with EtOAc (3 x 10.0 mL). The combined organic phases were washed with brine, dried over MgSO₄ and concentrated *in vacuo*. The residue was used in the next step without further purification.

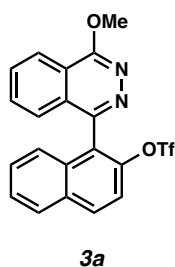
To a solution of crude mixture of **S2** (2.00 mmol, 1.00 equiv) and DMAP (4.00 mmol, 2.00 equiv) in CH₂Cl₂ (20.0 mL) was added Tf₂O (2.00 mmol, 1.00 equiv) dropwise at 0 °C. The reaction mixture was stirred at 23 °C until **S2** was fully consumed by TLC analysis. The reaction mixture was diluted with CH₂Cl₂ (10.0 mL) and water (10.0 mL). The aqueous phase was extracted with CH₂Cl₂ (3 x 6.00 mL). The combined organic phases were washed with brine, dried over MgSO₄ and concentrated *in vacuo*. The residue was purified by flash column chromatography (1:4 EtOAc:hexanes) on silica gel to give the corresponding triflate substrates.

General Procedure of PINAP from Triflates via Dynamic Kinetic Asymmetric Transformation



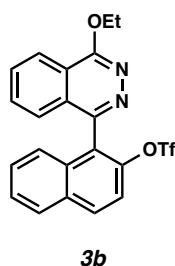
This reaction was performed in a nitrogen-filled glovebox. $\text{Pd}[(o\text{-tol})_3\text{P}]_2$ (10.8 mg, 0.0151 mmol) and (S, R_{Fc}) -Josiphos (13.7 mg, 0.0226 mmol) in dioxane (0.302 mL) were pre-stirred in a vial until all the solids were dissolved. To a solution of triflates **3** (0.211 mmol, 1.00 equiv) in dioxane (1.06 mL) was added DMAP (0.844 mmol, 4.00 equiv) and pre-stirred $\text{Pd}[(o\text{-tol})_3\text{P}]_2$ and (S, R_{Fc}) -Josiphos (0.05 M in dioxane; 0.00211 mmol, 0.01 equiv) at 23 °C. The mixture was placed in a reaction well preheated to 60 °C. A solution of Ph_2PH (1.00 M in dioxane; 0.317 mmol, 1.50 equiv) was added to the reaction mixture in 20 μL portions every 30 minutes manually. After completion of the addition (8 hours), the reaction was stirred for further 7 hours at which point complete consumption of the starting material was observed. The reaction was cooled, removed from the glovebox and diluted with EtOAc (1.50 mL) and water (2.00 mL). The aqueous phase was extracted with EtOAc (3 x 1.50 mL). The combined organic phases were washed with brine, dried with MgSO_4 and concentrated. The crude material was purified by flash column chromatography on silica gel to afford the corresponding PINAP **4**.

Spectroscopic Data

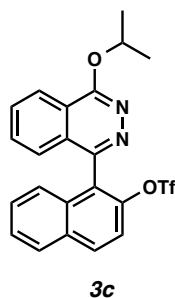


452 mg, 52% (2-step yield); R_f = 0.29 (1:4 EtOAc:hexanes); ^1H NMR (500 MHz, CDCl_3) δ 8.35 (dd, J = 8.2, 1.0 Hz, 1H), 8.12 (d, J = 9.1 Hz, 1H), 8.00 (d, J = 8.2 Hz,

1H), 7.87 (ddd, $J = 8.2, 7.0, 1.1$ Hz, 1H), 7.70 (ddd, $J = 8.4, 7.1, 1.3$ Hz, 1H), 7.62 (d, $J = 9.1$ Hz, 1H), 7.58 (ddd, $J = 8.2, 6.8, 1.3$ Hz, 1H), 7.42 (ddd, $J = 8.2, 6.7, 1.3$ Hz, 1H), 7.36 (d, $J = 8.5$ Hz, 1H), 7.31 (dt, $J = 8.4, 1.0$ Hz, 1H), 4.41 (s, 3H); ^{13}C NMR (126 MHz, CDCl_3) δ 160.9, 151.1, 145.6, 133.6, 132.7, 132.6, 132.4, 131.9, 129.2, 128.4, 128.1, 127.4, 126.7, 126.6, 125.7, 123.5, 119.9, 119.6, 117.1, 55.3; IR (Neat Film NaCl) 3063, 2945, 1668, 1581, 1541, 1496, 1457, 1420, 1364, 1247, 1213, 1139, 952, 834 cm^{-1} ; HRMS (MM: ESI-APCI+) m/z calc'd for $\text{C}_{20}\text{H}_{14}\text{F}_3\text{N}_2\text{O}_4\text{S}$ $[\text{M}+\text{H}]^+$: 435.0621; found: 435.0625.

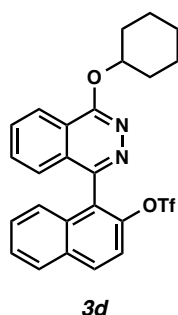


430 mg, 48% (2-step yield); $R_f = 0.33$ (1:4 EtOAc:hexanes); ^1H NMR (500 MHz, Chloroform- d) δ 8.38 (dd, $J = 8.2, 1.2$ Hz, 1H), 8.12 (d, $J = 9.1$ Hz, 1H), 7.99 (d, $J = 8.3$ Hz, 1H), 7.87 (ddd, $J = 8.2, 6.9, 1.2$ Hz, 1H), 7.69 (ddd, $J = 8.4, 7.1, 1.3$ Hz, 1H), 7.61 (d, $J = 9.0$ Hz, 1H), 7.57 (ddd, $J = 8.2, 6.7, 1.4$ Hz, 1H), 7.42 (ddd, $J = 8.1, 6.7, 1.3$ Hz, 1H), 7.37 (d, $J = 8.5$ Hz, 1H), 7.29 (dd, $J = 8.3, 1.0$ Hz, 1H), 4.88 (qd, $J = 7.0, 3.9$ Hz, 2H), 1.62 (td, $J = 7.1, 1.6$ Hz, 3H); ^{13}C NMR (126 MHz, CDCl_3) δ 160.6, 150.7, 145.6, 133.6, 132.7, 132.5, 132.3, 131.8, 129.2, 128.4, 128.1, 127.4, 126.7, 126.6, 125.6, 123.6, 120.0, 119.6, 117.1, 63.9, 14.7; IR (Neat Film NaCl) 3066, 2984, 1582, 1540, 1495, 1422, 1381, 1342, 1214, 1140, 1073, 1024, 956, 941, 834 cm^{-1} ; HRMS (MM: ESI-APCI+) m/z calc'd for $\text{C}_{21}\text{H}_{16}\text{F}_3\text{N}_2\text{O}_4\text{S}$ $[\text{M}+\text{H}]^+$: 449.0777; found: 449.0780.

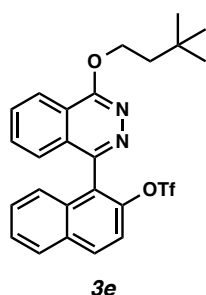


342 mg, 37% (2-step yield); $R_f = 0.35$ (1:4 EtOAc:hexanes); ^1H NMR (500 MHz, CDCl_3) δ 8.37 (dd, $J = 8.2, 1.0$ Hz, 1H), 8.13 (d, $J = 9.1$ Hz, 1H), 8.00 (d, $J = 8.3$ Hz,

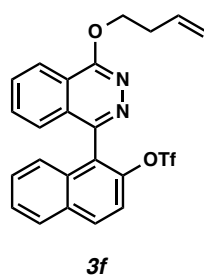
1H), 7.87 (ddd, $J = 8.2, 7.0, 1.2$ Hz, 1H), 7.69 (ddd, $J = 8.4, 7.1, 1.3$ Hz, 1H), 7.62 (d, $J = 9.1$ Hz, 1H), 7.58 (ddd, $J = 8.2, 6.7, 1.3$ Hz, 1H), 7.43 (ddd, $J = 8.1, 6.7, 1.3$ Hz, 1H), 7.40 – 7.35 (m, 1H), 7.30 (dt, $J = 8.3, 0.9$ Hz, 1H), 5.94 (hept, $J = 6.2$ Hz, 1H), 1.60 (d, $J = 6.1$ Hz, 3H), 1.58 (d, $J = 6.2$ Hz, 3H); ^{13}C NMR (126 MHz, CDCl_3) δ 160.3, 150.2, 145.7, 133.5, 132.7, 132.5, 132.4, 132.0, 129.3, 128.5, 128.2, 127.4, 126.6, 126.4, 125.7, 123.7, 120.4, 119.7, 71.1, 22.3, 22.1; IR (Neat Film NaCl) 2981, 1582, 1493, 1417, 1322, 1212, 1140, 1106, 958, 942, 833, 748 cm^{-1} ; HRMS (MM: ESI-APCI+) m/z calc'd for $\text{C}_{22}\text{H}_{17}\text{F}_3\text{N}_2\text{O}_4\text{S}$ $[\text{M}+\text{H}]^+$: 463.0934; found: 463.0964.



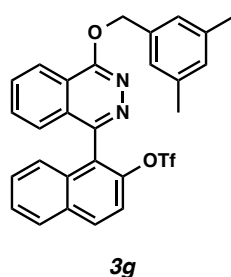
362 mg, 36% (2-step yield); $R_f = 0.43$ (1:4 EtOAc:hexanes); ^1H NMR (500 MHz, CDCl_3) δ 8.37 (d, $J = 8.2$ Hz, 1H), 8.11 (dd, $J = 9.1, 1.6$ Hz, 1H), 7.99 (d, $J = 8.3$ Hz, 1H), 7.85 (ddd, $J = 8.4, 7.1, 1.3$ Hz, 1H), 7.67 (td, $J = 7.7, 7.0, 1.4$ Hz, 1H), 7.61 (dd, $J = 9.1, 2.0$ Hz, 1H), 7.57 (ddd, $J = 8.2, 6.2, 1.9$ Hz, 1H), 7.44 – 7.37 (m, 2H), 7.27 (d, $J = 8.3$ Hz, 1H), 5.72 (tt, $J = 8.7, 3.7$ Hz, 1H), 2.25 (d, $J = 12.3$ Hz, 2H), 1.90 (ddt, $J = 13.9, 8.9, 4.5$ Hz, 2H), 1.85 – 1.74 (m, 2H), 1.66 (td, $J = 8.5, 7.7, 4.2$ Hz, 1H), 1.57 (ddt, $J = 13.5, 10.2, 3.4$ Hz, 2H), 1.45 (ddt, $J = 13.3, 10.1, 3.5$ Hz, 1H); ^{13}C NMR (126 MHz, CDCl_3) δ 160.2, 150.3, 145.6, 133.6, 132.7, 132.3, 132.1, 131.7, 129.3, 128.4, 128.1, 127.3, 126.9, 126.7, 125.5, 123.7, 120.3, 119.7, 75.4, 32.0, 31.8, 25.9, 24.1; IR (Neat Film NaCl) 2937, 2858, 1582, 1537, 1493, 1418, 1362, 1342, 1214, 1140, 959, 944, 834 cm^{-1} ; HRMS (MM: ESI-APCI+) m/z calc'd for $\text{C}_{25}\text{H}_{22}\text{F}_3\text{N}_2\text{O}_4\text{S}$ $[\text{M}+\text{H}]^+$: 503.1247; found: 503.1248.



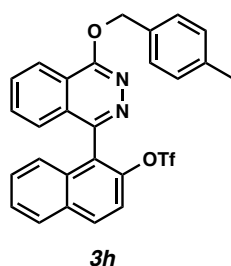
494 mg, 49% (2-step yield); R_f = 0.33 (1:4 EtOAc:hexanes); ^1H NMR (500 MHz, CDCl_3) δ 8.35 (d, J = 8.2 Hz, 1H), 8.12 (d, J = 9.1 Hz, 1H), 8.00 (d, J = 8.2 Hz, 1H), 7.86 (ddd, J = 8.2, 7.0, 1.1 Hz, 1H), 7.69 (ddd, J = 8.3, 7.1, 1.2 Hz, 1H), 7.62 (d, J = 9.1 Hz, 1H), 7.57 (ddd, J = 8.2, 6.6, 1.3 Hz, 1H), 7.42 (ddd, J = 8.1, 6.6, 1.2 Hz, 1H), 7.39 – 7.35 (m, 1H), 7.29 (d, J = 8.3 Hz, 1H), 4.88 (t, J = 7.2 Hz, 2H), 1.96 (t, J = 7.2 Hz, 2H), 1.10 (s, 9H); ^{13}C NMR (126 MHz, CDCl_3) δ 160.7, 150.7, 145.6, 133.6, 132.7, 132.4, 132.3, 131.8, 129.2, 128.4, 128.1, 127.4, 126.6, 125.6, 123.5, 120.0, 119.6, 117.1, 110.2, 65.8, 42.5, 30.0; IR (Neat Film NaCl) 2958, 1582, 1538, 1495, 1422, 1360, 1214, 1141, 1073, 953, 834, 620 cm^{-1} ; HRMS (MM: ESI-APCI+) m/z calc'd for $\text{C}_{25}\text{H}_{23}\text{F}_3\text{N}_2\text{O}_4\text{S}$ $[\text{M}+\text{H}]^+$: 505.1403; found: 505.1436.



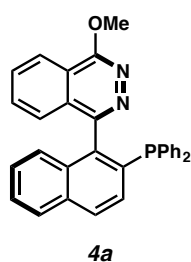
493 mg, 52% (2-step yield); R_f = 0.33 (1:4 EtOAc:hexanes); ^1H NMR (500 MHz, CDCl_3) δ 8.38 – 8.34 (m, 1H), 8.12 (d, J = 9.1 Hz, 1H), 8.00 (d, J = 8.2 Hz, 1H), 7.87 (ddd, J = 8.3, 7.0, 1.1 Hz, 1H), 7.69 (ddd, J = 8.4, 7.0, 1.3 Hz, 1H), 7.61 (d, J = 9.1 Hz, 1H), 7.57 (ddd, J = 8.2, 6.7, 1.3 Hz, 1H), 7.42 (ddd, J = 8.1, 6.7, 1.2 Hz, 1H), 7.38 – 7.35 (m, 1H), 7.30 (dd, J = 8.3, 1.1 Hz, 1H), 6.04 (ddt, J = 17.1, 10.3, 6.7 Hz, 1H), 5.28 (dq, J = 17.2, 1.7 Hz, 1H), 5.17 (dq, J = 10.3, 1.4 Hz, 1H), 4.94 – 4.83 (m, 2H), 2.78 (dddd, J = 8.1, 6.6, 5.2, 1.4 Hz, 2H); ^{13}C NMR (126 MHz, CDCl_3) δ 160.6, 150.9, 145.6, 134.6, 133.6, 132.7, 132.5, 132.3, 131.8, 129.2, 128.4, 128.1, 127.4, 126.7, 126.6, 125.6, 123.5, 119.9, 119.6, 117.3, 117.1, 67.0, 33.6; IR (Neat Film NaCl) 1581, 1538, 1494, 1421, 1349, 1213, 1139, 954, 833 cm^{-1} ; HRMS (MM: ESI-APCI+) m/z calc'd for $\text{C}_{23}\text{H}_{18}\text{F}_3\text{N}_2\text{O}_4\text{S}$ $[\text{M}+\text{H}]^+$: 475.0934; found: 475.0956.



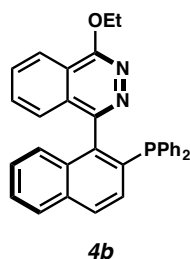
377 mg, 35% (2-step yield); R_f = 0.38 (1:4 EtOAc:hexanes); ^1H NMR (500 MHz, CDCl_3) δ 8.40 (dt, J = 8.2, 1.0 Hz, 1H), 8.13 (d, J = 9.1 Hz, 1H), 8.00 (dt, J = 8.3, 0.9 Hz, 1H), 7.86 (ddd, J = 8.3, 7.1, 1.2 Hz, 1H), 7.70 (ddd, J = 8.3, 7.1, 1.2 Hz, 1H), 7.63 (d, J = 9.1 Hz, 1H), 7.58 (ddd, J = 8.2, 6.6, 1.3 Hz, 1H), 7.43 (ddd, J = 7.9, 6.6, 1.3 Hz, 1H), 7.39 (dd, J = 8.6, 1.2 Hz, 1H), 7.31 (dt, J = 8.4, 1.0 Hz, 1H), 7.04 (s, 1H), 5.80 (d, J = 2.3 Hz, 2H), 2.39 (s, 6H); ^{13}C NMR (126 MHz, CDCl_3) δ 160.6, 151.1, 145.6, 138.3, 136.7, 133.6, 132.7, 132.6, 132.3, 131.8, 130.1, 129.3, 128.4, 128.1, 127.4, 126.7, 126.6, 126.6, 125.6, 123.7, 119.9, 119.7, 117.1, 69.8, 21.4; IR (Neat Film NaCl) 2917, 1581, 1494, 1418, 1343, 1213, 1140, 956, 835 cm^{-1} ; HRMS (MM: ESI-APCI+) m/z calc'd for $\text{C}_{28}\text{H}_{22}\text{F}_3\text{N}_2\text{O}_4\text{S}$ $[\text{M}+\text{H}]^+$: 539.1247; found: 539.1252.



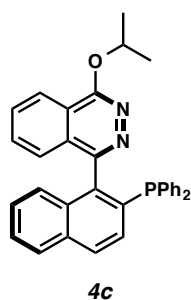
713 mg, 68% (2-step yield); R_f = 0.37 (1:4 EtOAc:hexanes); ^1H NMR (500 MHz, CDCl_3) δ 8.38 (dt, J = 8.2, 1.0 Hz, 1H), 8.13 (d, J = 9.1 Hz, 1H), 8.01 (dt, J = 8.3, 0.9 Hz, 1H), 7.85 (ddd, J = 8.3, 7.0, 1.2 Hz, 1H), 7.70 (ddd, J = 8.4, 7.1, 1.3 Hz, 1H), 7.62 (dd, J = 9.1, 1.1 Hz, 1H), 7.58 (ddd, J = 8.1, 6.8, 1.3 Hz, 1H), 7.57 – 7.52 (m, 2H), 7.43 (ddd, J = 8.1, 6.8, 1.3 Hz, 1H), 7.37 (dt, J = 8.6, 1.0 Hz, 1H), 7.31 (dt, J = 8.3, 1.0 Hz, 1H), 7.27 (d, J = 7.2 Hz, 2H), 5.81 (d, J = 7.1 Hz, 2H), 2.41 (s, 3H); ^{13}C NMR (126 MHz, CDCl_3) δ 160.5, 151.1, 145.5, 138.3, 133.7, 133.5, 132.6, 132.3, 131.9, 129.4, 129.2, 128.90, 128.4, 128.1, 127.4, 126.7, 126.6, 125.6, 123.6, 119.8, 119.6, 119.6, 117.0, 69.6, 21.4; IR (Neat Film NaCl) 3058, 1581, 1538, 1494, 1422, 1342, 1248, 1266, 1140, 1072, 953, 834, 776 cm^{-1} ; HRMS (MM: ESI-APCI+) m/z calc'd for $\text{C}_{27}\text{H}_{19}\text{F}_3\text{N}_2\text{O}_4\text{S}$ $[\text{M}+\text{H}]^+$: 525.1090; found: 525.1096.



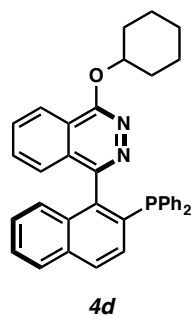
79 mg, 80% yield, 96% ee; $[\alpha]_{\text{D}}^{25} +124.5$ (c 0.51, CHCl_3); $R_f = 0.42$ (1:2 EtOAc:hexanes); ^1H NMR (500 MHz, CDCl_3) δ 8.27 (dt, $J = 8.3, 1.0$ Hz, 1H), 7.91 (dd, $J = 8.6, 0.8$ Hz, 1H), 7.90 – 7.87 (m, 1H), 7.74 (ddd, $J = 8.2, 7.1, 1.1$ Hz, 1H), 7.49 (ddd, $J = 8.2, 6.8, 1.2$ Hz, 1H), 7.46 – 7.42 (m, 2H), 7.29 (ddd, $J = 8.2, 6.8, 1.3$ Hz, 1H), 7.27 – 7.14 (m, 11H), 7.07 (dt, $J = 8.2, 0.9$ Hz, 1H), 4.36 (s, 3H); (Due to the C–P coupling, the ^{13}C NMR contained extra peaks. See the attached spectrum); ^{13}C NMR (126 MHz, CDCl_3) δ 160.4, 156.8, 156.8, 141.5, 141.3, 137.5, 137.4, 137.3, 137.2, 136.0, 135.9, 134.0, 133.9, 133.7, 133.3, 133.2, 133.1, 131.8, 131.7, 130.3, 130.0, 129.9, 129.3, 128.7, 128.4, 128.4, 128.2, 128.1, 127.1, 126.9, 126.7, 126.7, 126.1, 123.2, 119.6, 55.1; ^{31}P NMR (121 MHz, CDCl_3) δ -14.3; IR (Neat Film NaCl) 3053, 1582, 1538, 1495, 1456, 1362, 1112, 1018, 744 cm^{-1} ; HRMS (MM: ESI-APCI+) m/z calc'd for $\text{C}_{31}\text{H}_{23}\text{N}_2\text{OP}$ $[\text{M}+\text{H}]^+$: 471.1621; found: 471.1674; SFC conditions: 35% IPA, 2.5mL/min, Chiralcel OD-H column, t_R (min): major = 5.81, minor = 5.16.



75 mg, 73% yield, 94% ee; $[\alpha]_{\text{D}}^{25} +150.3$ (c 0.47, CHCl_3); $R_f = 0.54$ (1:2 EtOAc:hexanes); ^1H NMR (500 MHz, CDCl_3) δ 8.30 (dd, $J = 8.2, 1.0$ Hz, 1H), 7.91 (d, $J = 8.6$ Hz, 1H), 7.89 (d, $J = 7.8$ Hz, 1H), 7.74 (ddd, $J = 8.2, 7.1, 1.1$ Hz, 1H), 7.49 (ddd, $J = 8.1, 6.8, 1.3$ Hz, 1H), 7.47 – 7.41 (m, 2H), 7.29 (ddd, $J = 8.2, 6.8, 1.3$ Hz, 1H), 7.27 – 7.13 (m, 11H), 7.07 (d, $J = 8.2$ Hz, 1H), 4.83 (qt, $J = 7.1, 2.1$ Hz, 2H), 1.60 (t, $J = 7.1$ Hz, 3H); (Due to the C–P coupling, the ^{13}C NMR contained extra peaks. See the attached spectrum); ^{13}C NMR (126 MHz, CDCl_3) δ 160.2, 156.5, 156.5, 141.6, 141.3, 137.6, 137.5, 137.3, 137.2, 136.0, 135.9, 134.0, 133.8, 133.7, 133.3, 133.2, 133.1, 131.8, 131.5, 130.3, 130.0, 129.9, 129.2, 128.6, 128.4, 128.4, 128.2, 128.1, 127.1, 126.8, 126.8, 126.7, 126.1, 123.3, 119.6, 63.5, 14.9; ^{31}P NMR (121 MHz, CDCl_3) δ -14.2; IR (Neat Film NaCl) 3053, 2980, 1583, 1538, 1494, 1418, 1378, 1342, 1165, 1110, 1023, 818, 744 cm^{-1} ; HRMS (MM: ESI-APCI+) m/z calc'd for $\text{C}_{32}\text{H}_{25}\text{N}_2\text{OP}$ $[\text{M}+\text{H}]^+$: 485.1777; found: 485.1800; SFC conditions: 30% MeOH, 2.5mL/min, Chiralpak AD-H column, t_R (min): major = 2.60, minor = 3.09.

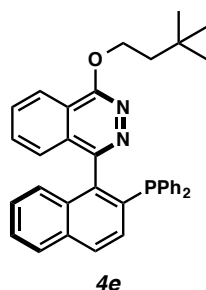


74 mg, 70% yield, 82% ee; $[\alpha]_{\text{D}}^{25} +131.7$ (*c* 0.57, CHCl_3); $R_f = 0.54$ (1:2 EtOAc:hexanes); ^1H NMR (500 MHz, CDCl_3) δ 8.29 (dt, $J = 8.1, 1.0$ Hz, 1H), 7.90 (t, $J = 9.0$ Hz, 2H), 7.73 (ddd, $J = 8.2, 7.1, 1.1$ Hz, 1H), 7.52 – 7.42 (m, 3H), 7.30 (ddd, $J = 8.4, 6.6, 1.2$ Hz, 1H), 7.28 – 7.15 (m, 11H), 7.08 (dt, $J = 8.1, 0.9$ Hz, 1H), 5.88 (p, $J = 6.2$ Hz, 1H), 1.58 (d, $J = 5.7$ Hz, 3H), 1.56 (d, $J = 5.7$ Hz, 3H); (Due to the C–P coupling, the ^{13}C NMR contained extra peaks. See the attached spectrum); ^{13}C NMR (126 MHz, CDCl_3) δ 159.7, 156.0, 156.0, 141.6, 141.3, 137.6, 137.5, 137.3, 137.2, 136.0, 135.9, 134.0, 133.8, 133.7, 133.4, 133.2, 133.2, 133.1, 131.7, 131.4, 130.2, 130.0, 129.2, 128.6, 128.4, 128.4, 128.4, 128.4, 128.2, 128.1, 127.0, 126.8, 126.0, 123.4, 120.0, 70.1, 22.4, 22.3; ^{31}P NMR (121 MHz, CDCl_3) δ -13.9; IR (Neat Film NaCl) 3053, 2978, 1584, 1489, 1413, 1383, 1315, 1106, 744 cm^{-1} ; HRMS (MM: ESI-APCI+) *m/z* calc'd for $\text{C}_{33}\text{H}_{27}\text{N}_2\text{OP}$ $[\text{M}+\text{H}]^+$: 499.1934; found: 499.1960; SFC conditions: 40% MeOH, 2.5mL/min, Chiralpak AD-H column, t_R (min): major = 1.94, minor = 2.22.

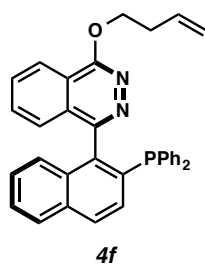


76 mg, 67% yield, 95% ee; $[\alpha]_{\text{D}}^{25} +91.0$ (*c* 0.57, CHCl_3); $R_f = 0.61$ (1:2 EtOAc:hexanes); ^1H NMR (500 MHz, CDCl_3) δ 8.29 (dt, $J = 8.2, 1.0$ Hz, 1H), 7.89 (ddd, $J = 9.4, 8.2, 0.8$ Hz, 2H), 7.73 (ddd, $J = 8.3, 7.0, 1.2$ Hz, 1H), 7.49 (ddd, $J = 8.1, 6.7, 1.3$ Hz, 1H), 7.46 – 7.41 (m, 2H), 7.29 (ddd, $J = 8.2, 6.7, 1.3$ Hz, 1H), 7.26 – 7.14 (m, 11H), 7.07 (dt, $J = 8.3, 0.9$ Hz, 1H), 5.67 (tt, $J = 8.9, 3.8$ Hz, 1H), 2.30 – 2.18 (m, 2H), 1.88 (ddtd, $J = 9.3, 7.9, 5.4, 4.0$ Hz, 2H), 1.76 (qdt, $J = 9.9, 6.1, 3.7$ Hz, 2H),

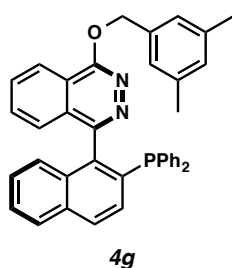
1.64 (tt, $J = 9.1, 4.9$ Hz, 1H), 1.54 (qq, $J = 10.2, 3.5$ Hz, 2H), 1.42 (ddt, $J = 13.4, 10.2, 3.5$ Hz, 1H); (Due to the C–P coupling, the ^{13}C NMR contained extra peaks. See the attached spectrum); ^{13}C NMR (126 MHz, CDCl_3) δ 159.6, 156.0, 155.9, 141.6, 141.4, 137.6, 137.5, 137.3, 137.2, 136.0, 135.9, 134.0, 133.9, 133.7, 133.4, 133.2, 131.7, 131.4, 130.3, 130.0, 129.2, 128.6, 128.4, 128.4, 128.4, 128.4, 128.2, 128.1, 127.0, 126.8, 126.8, 126.8, 126.0, 123.4, 120.0, 74.8, 32.0, 31.9, 25.9, 24.1, 24.0; ^{31}P NMR (121 MHz, CDCl_3) δ -13.9; IR (Neat Film NaCl) 3051, 2933, 2856, 1582, 1538, 1490, 1432, 1413, 1361, 1341, 1311, 1263, 1215, 1165, 1113, 1068, 1046, 1019, 744 cm^{-1} ; HRMS (MM: ESI-APCI+) m/z calc'd for $\text{C}_{36}\text{H}_{31}\text{N}_2\text{OP}$ $[\text{M}+\text{H}]^+$: 539.2247; found: 539.2268; SFC conditions: 25% MeOH, 2.5mL/min, Chiralcel OD-H column, t_R (min): major = 7.58, minor = 8.13.



65 mg, 57% yield, 95% ee; $[\alpha]_{\text{D}}^{25} +101.2$ (c 0.42, CHCl_3); $R_f = 0.62$ (1:2 EtOAc:hexanes); ^1H NMR (500 MHz, CDCl_3) δ 8.27 (dt, $J = 8.2, 1.0$ Hz, 1H), 7.91 (d, $J = 8.5$ Hz, 1H), 7.89 (d, $J = 8.4$ Hz, 1H), 7.73 (ddt, $J = 8.7, 7.1, 1.4$ Hz, 1H), 7.49 (ddt, $J = 8.5, 6.9, 1.5$ Hz, 1H), 7.46 – 7.40 (m, 2H), 7.29 (ddd, $J = 8.2, 6.7, 1.3$ Hz, 1H), 7.27 – 7.14 (m, 11H), 7.06 (d, $J = 8.2$ Hz, 1H), 4.87 – 4.77 (m, 2H), 1.94 (td, $J = 7.1, 1.9$ Hz, 2H), 1.09 (s, 9H); (Due to the C–P coupling, the ^{13}C NMR contained extra peaks. See the attached spectrum); ^{13}C NMR (126 MHz, CDCl_3) δ 160.3, 156.5, 156.4, 141.6, 141.3, 137.6, 137.5, 137.3, 137.2, 136.0, 135.9, 134.0, 133.9, 133.7, 133.3, 133.2, 133.1, 131.7, 131.6, 130.3, 129.9, 129.2, 128.7, 128.4, 128.4, 128.2, 128.1, 127.1, 126.8, 126.8, 126.7, 126.1, 123.2, 119.7, 77.2, 65.4, 42.5, 30.1; ^{31}P NMR (121 MHz, CDCl_3) δ -14.2; IR (Neat Film NaCl) 3051, 2956, 1418, 1386, 1114, 743 cm^{-1} ; HRMS (MM: ESI-APCI+) m/z calc'd for $\text{C}_{36}\text{H}_{33}\text{N}_2\text{OP}$ $[\text{M}+\text{H}]^+$: 541.2403; found: 541.2409; SFC conditions: 35% IPA, 2.5mL/min, Chiralcel OD-H column, t_R (min): major = 4.33, minor = 4.84.

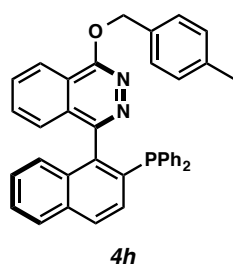


72 mg, 67% yield, 94% ee; $[\alpha]_{\text{D}}^{25} +117.9$ (c 0.52, CHCl_3); $R_f = 0.62$ (1:2 EtOAc:hexanes); ^1H NMR (500 MHz, CDCl_3) δ 8.28 (d, $J = 8.2$ Hz, 1H), 7.91 (d, $J = 8.6$ Hz, 1H), 7.89 (d, $J = 8.2$ Hz, 1H), 7.75 (ddd, $J = 8.3, 7.1, 1.2$ Hz, 1H), 7.52 – 7.41 (m, 3H), 7.30 (ddd, $J = 8.3, 6.8, 1.3$ Hz, 1H), 7.26 – 7.14 (m, 11H), 7.08 (dt, $J = 8.4, 0.9$ Hz, 1H), 6.04 (ddt, $J = 17.0, 10.3, 6.7$ Hz, 1H), 5.28 (dq, $J = 17.2, 1.7$ Hz, 1H), 5.17 (dt, $J = 10.5, 1.6$ Hz, 1H), 4.88 – 4.77 (m, 2H), 2.75 (qt, $J = 6.8, 1.3$ Hz, 2H); (Due to the C–P coupling, the ^{13}C NMR contained extra peaks. See the attached spectrum); ^{13}C NMR (126 MHz, CDCl_3) δ 160.1, 156.7, 156.6, 141.5, 141.2, 137.6, 137.5, 137.2, 137.2, 136.0, 135.9, 134.9, 134.0, 133.8, 133.7, 133.3, 133.2, 133.2, 133.1, 131.9, 131.6, 130.3, 130.0, 130.0, 129.3, 128.7, 128.4, 128.4, 128.2, 128.1, 127.1, 126.9, 126.7, 126.7, 126.1, 123.2, 119.6, 117.2, 77.2, 66.7, 33.7; ^{31}P NMR (121 MHz, CDCl_3) δ -14.2; IR (Neat Film NaCl) 2096, 1642, 1417, 1348, 1113, 1020, 818, 744 cm^{-1} ; HRMS (MM: ESI-APCI+) m/z calc'd for $\text{C}_{34}\text{H}_{27}\text{N}_2\text{OP}$ $[\text{M}+\text{H}]^+$: 511.1934; found: 511.1954; SFC conditions: 35% MeOH, 2.5mL/min, Chiralpak AD-H column, t_R (min): major = 3.00, minor = 3.66.



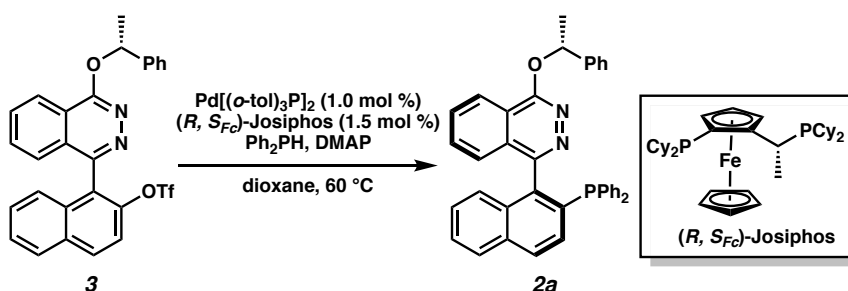
73 mg, 60% yield, 94% ee; $[\alpha]_{\text{D}}^{25} +40.1$ (c 0.31, CHCl_3); $R_f = 0.56$ (1:2 EtOAc:hexanes); ^1H NMR (300 MHz, CDCl_3) δ 8.32 (d, $J = 8.2$ Hz, 1H), 7.91 (t, $J = 7.8$ Hz, 2H), 7.73 (ddd, $J = 8.3, 7.1, 1.3$ Hz, 1H), 7.54 – 7.41 (m, 3H), 7.35 – 7.13 (m, 14H), 7.12 – 7.03 (m, 2H), 5.74 (d, $J = 5.4$ Hz, 2H), 2.39 (s, 6H); (Due to the C–P coupling, the ^{13}C NMR contained extra peaks. See the attached spectrum); ^{13}C NMR (75 MHz, CDCl_3) δ 160.1, 156.9, 156.8, 141.5, 141.1, 138.3, 137.5, 137.4, 137.3, 137.1, 136.9, 136.0, 135.9, 134.1, 133.8, 133.7, 133.4, 133.2, 133.1, 133.1, 131.9,

131.6, 130.3, 130.0, 130.0, 129.3, 128.7, 128.5, 128.4, 128.4, 128.2, 128.1, 127.1, 126.9, 126.7, 126.7, 126.6, 126.6, 126.1, 123.4, 119.6, 69.5, 21.5; ^{31}P NMR (121 MHz, CDCl_3) δ -14.2; IR (Neat Film NaCl) 3052, 2917, 1608, 1582, 1538, 1492, 1480, 1432, 1414, 1386, 1345, 1264, 1165, 1113, 1019, 962, 851, 818, 777, 743 cm^{-1} ; HRMS (MM: ESI-APCI+) m/z calc'd for $\text{C}_{39}\text{H}_{31}\text{N}_2\text{OP}$ $[\text{M}+\text{H}]^+$: 575.2247; found: 575.2293; SFC conditions: 40% MeOH, 2.5mL/min, Chiralcel OD-H column, t_R (min): major = 5.77, minor = 6.82.



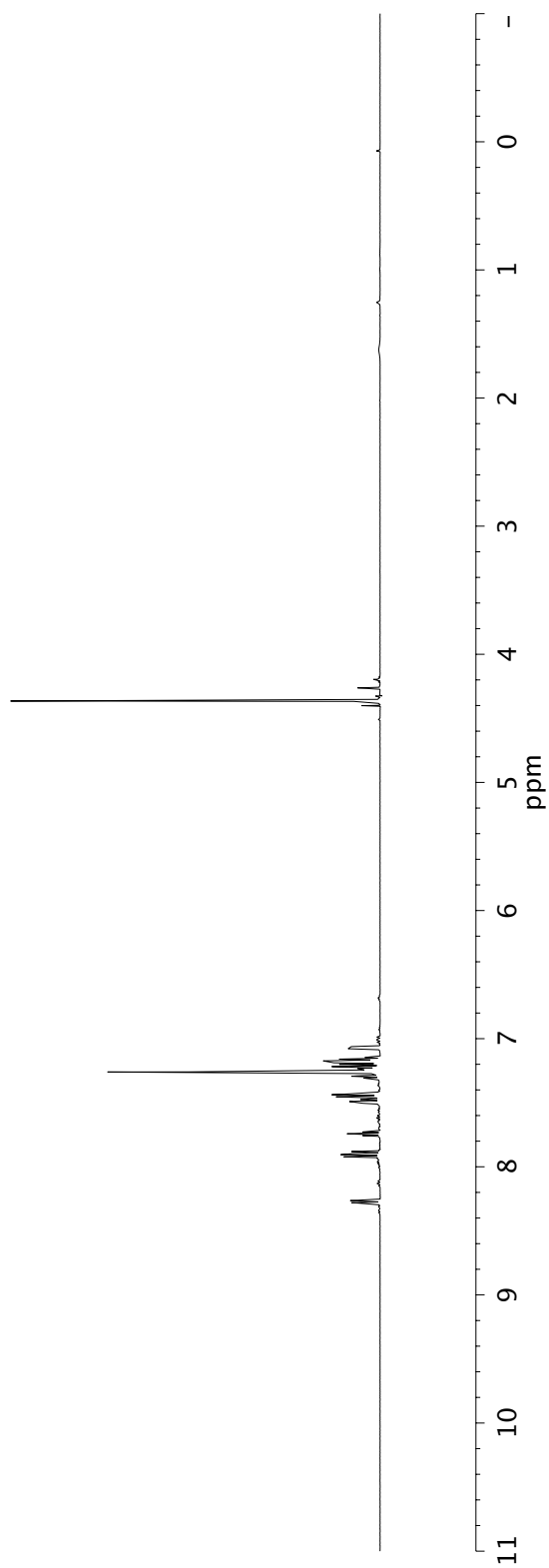
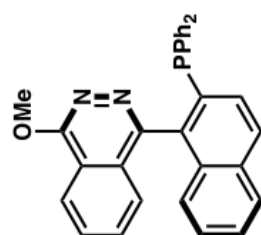
85 mg, 72% yield, 94% ee; $[\alpha]_{\text{D}}^{25}$ +118.5 (c 0.33, CHCl_3); R_f = 0.61 (1:2 EtOAc:hexanes); ^1H NMR (500 MHz, CDCl_3) δ 8.29 (dt, J = 8.2, 1.7 Hz, 1H), 7.91 (dd, J = 8.6, 2.5 Hz, 1H), 7.88 (dd, J = 8.2, 1.1 Hz, 1H), 7.71 (ddd, J = 8.2, 7.0, 1.2 Hz, 1H), 7.52 (dd, J = 8.2, 2.6 Hz, 2H), 7.48 (ddd, J = 8.2, 6.7, 1.3 Hz, 1H), 7.43 (dddd, J = 9.2, 6.0, 3.4, 1.8 Hz, 2H), 7.32 – 7.13 (m, 14H), 7.10 – 7.06 (m, 1H), 5.76 (q, J = 12.1 Hz, 2H), 2.40 (s, 3H); (Due to the C–P coupling, the ^{13}C NMR contained extra peaks. See the attached spectrum); ^{13}C NMR (126 MHz, CDCl_3) δ 160.1, 156.9, 156.8, 141.4, 141.2, 138.1, 137.5, 137.4, 137.2, 137.1, 136.0, 135.9, 134.0, 133.9, 133.7, 133.4, 133.2, 133.1, 131.9, 131.6, 130.3, 130.0, 130.0, 129.4, 129.4, 129.3, 128.9, 128.7, 128.4, 128.4, 128.4, 128.4, 128.3, 128.1, 127.1, 126.9, 126.7, 126.7, 126.1, 123.3, 119.6, 69.3, 21.4; ^{31}P NMR (121 MHz, CDCl_3) δ -14.2; IR (Neat Film NaCl) 3051, 1583, 1492, 1416, 1345, 1163, 1112, 1019, 744 cm^{-1} ; HRMS (MM: ESI-APCI+) m/z calc'd for $\text{C}_{38}\text{H}_{29}\text{N}_2\text{OP}$ $[\text{M}+\text{H}]^+$: 561.2090; found: 561.2090; SFC conditions: 40% MeOH, 2.5mL/min, Chiralpak AD-H column, t_R (min): major = 3.43, minor = 5.21.

A gram scale example

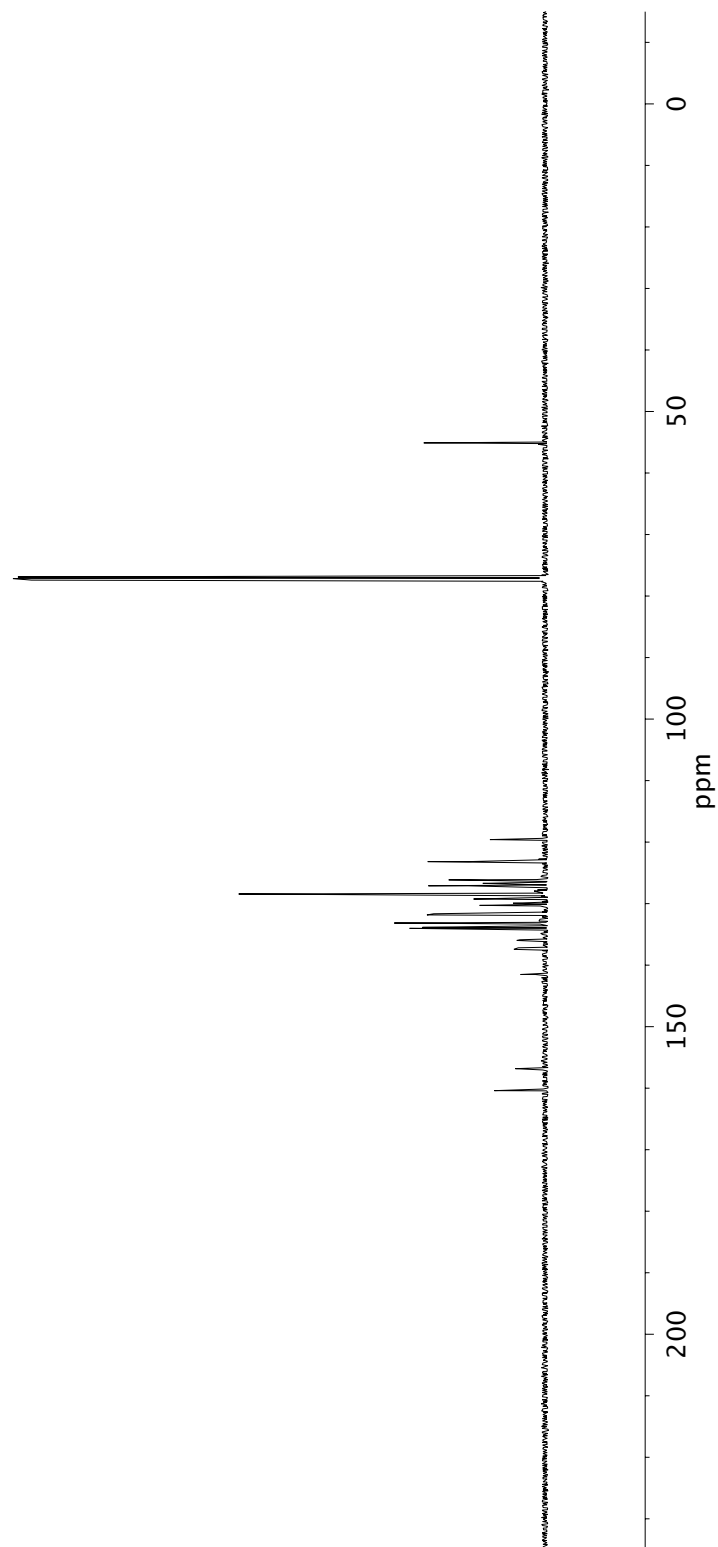
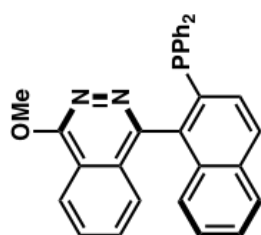


This reaction was performed in a nitrogen-filled glovebox. $\text{Pd}[(o\text{-tol})_3\text{P}]_2$ (17.9 mg, 0.0250 mmol) and $(R, S_{Fc})\text{-Josiphos}$ (22.7 mg, 0.0375 mmol) in dioxane (0.50 mL) were pre-stirred in a vial until all the solids were dissolved. To a solution of triflates **3** (1.24g, 2.36 mmol, 1.00 equiv) in dioxane (12.0 mL) was added DMAP (1.16g, 9.46 mmol, 4.00 equiv) and pre-stirred $\text{Pd}[(o\text{-tol})_3\text{P}]_2$ and $(R, S_{Fc})\text{-Josiphos}$ (0.05 M in dioxane; 0.47 mL, 0.0236 mmol, 0.01 equiv) at 23 °C. The mixture was placed in a reaction well preheated to 60 °C. A solution of Ph_2PH (1.00 M in dioxane; 3.54 mL, 3.54 mmol, 1.50 equiv) was added to the reaction mixture in 0.22 mL portions every 30 minutes manually. After completion of the addition (8 hours), the reaction was stirred for further 7 hours at which point complete consumption of the starting material was observed. The reaction was cooled, removed from the glovebox and diluted with EtOAc (15.0 mL) and water (20.0 mL). The aqueous phase was extracted with EtOAc (3 x 15.0 mL). The combined organic phases were washed with brine, dried with MgSO_4 and concentrated. The crude material was purified by flash column chromatography on silica gel (0.94 g, 71% yield, 88% de), and then recrystallized to afford PINAP **2a** (0.75 g, 57% yield, 99% de). All the other spectra data were identical to the reported data. (ref. 1)

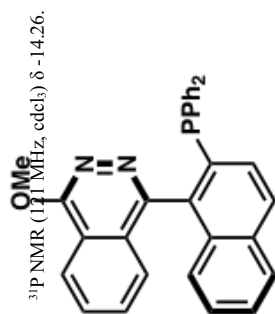
[1] a) T. F. Knöpfel, P. Aschwanden, T. Ichikawa, T. Watanabe, E. M. Carreira, *Angew. Chem., Int. Ed.* **2004**, *43*, 5971–5973. b) S. Fujimori, T. F. Knöpfel, P. Zarotti, T. Ichikawa, D. Boyall, E. M. Carreira, *Bull. Chem. Soc. Jpn.* **2007**, *80*, 1635–1657.



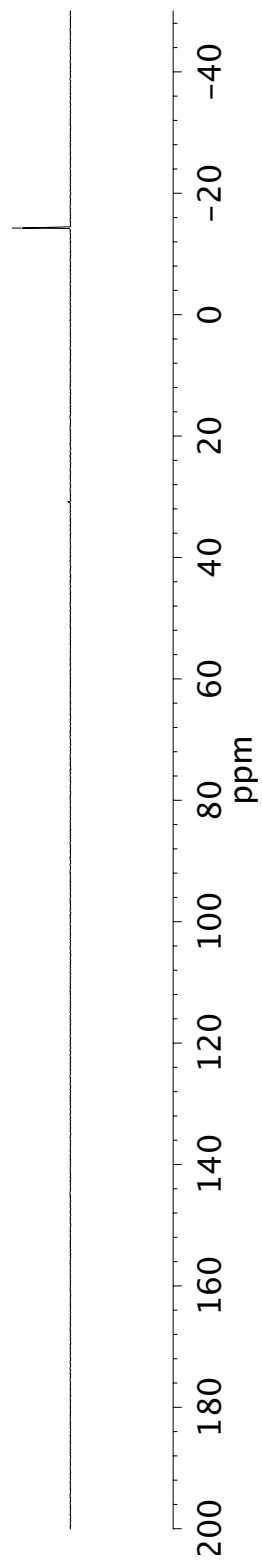
¹H NMR (500 MHz, CDCl₃) of compound **4a**.



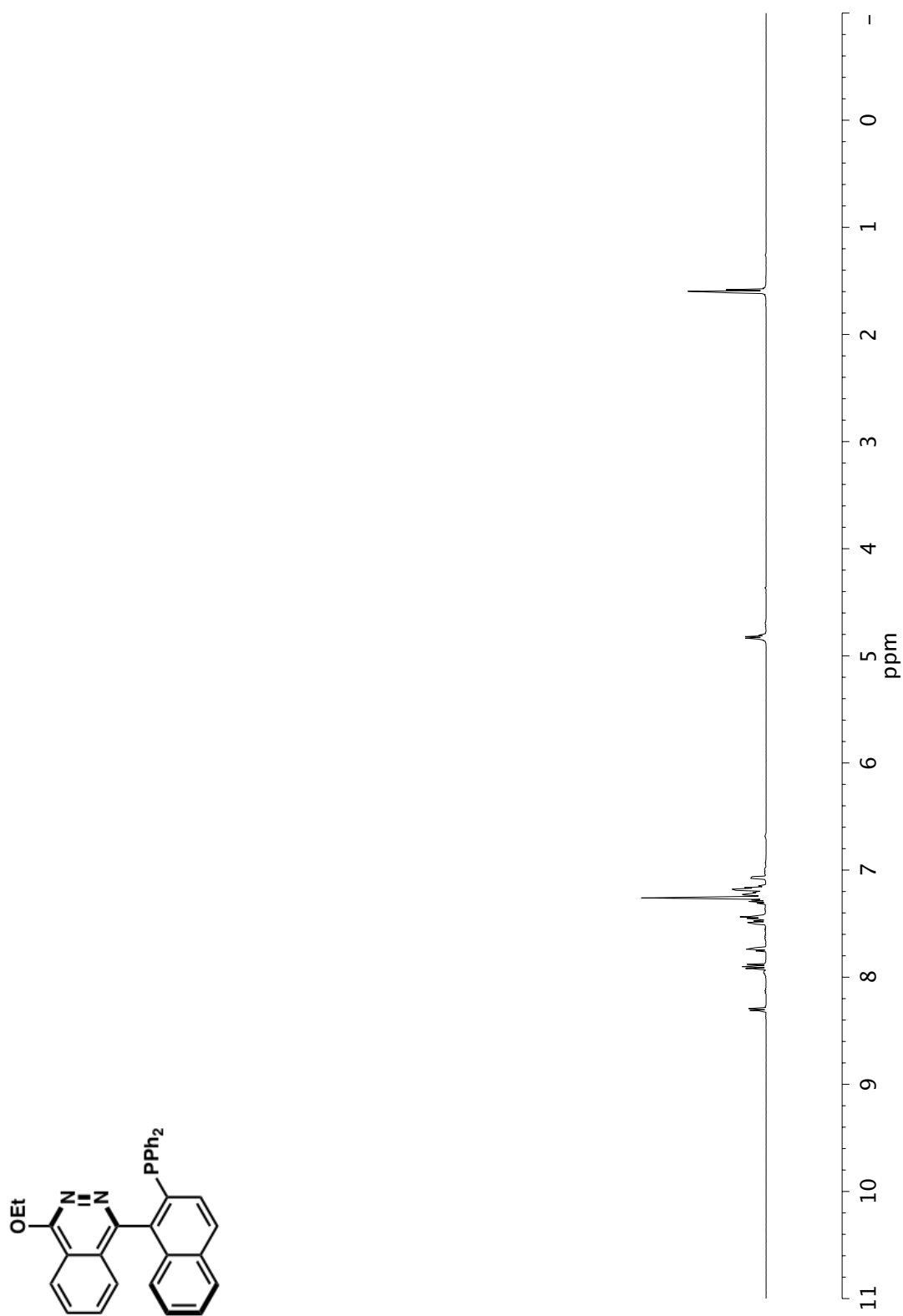
^{13}C NMR (126 MHz, CDCl_3) of compound 4a.

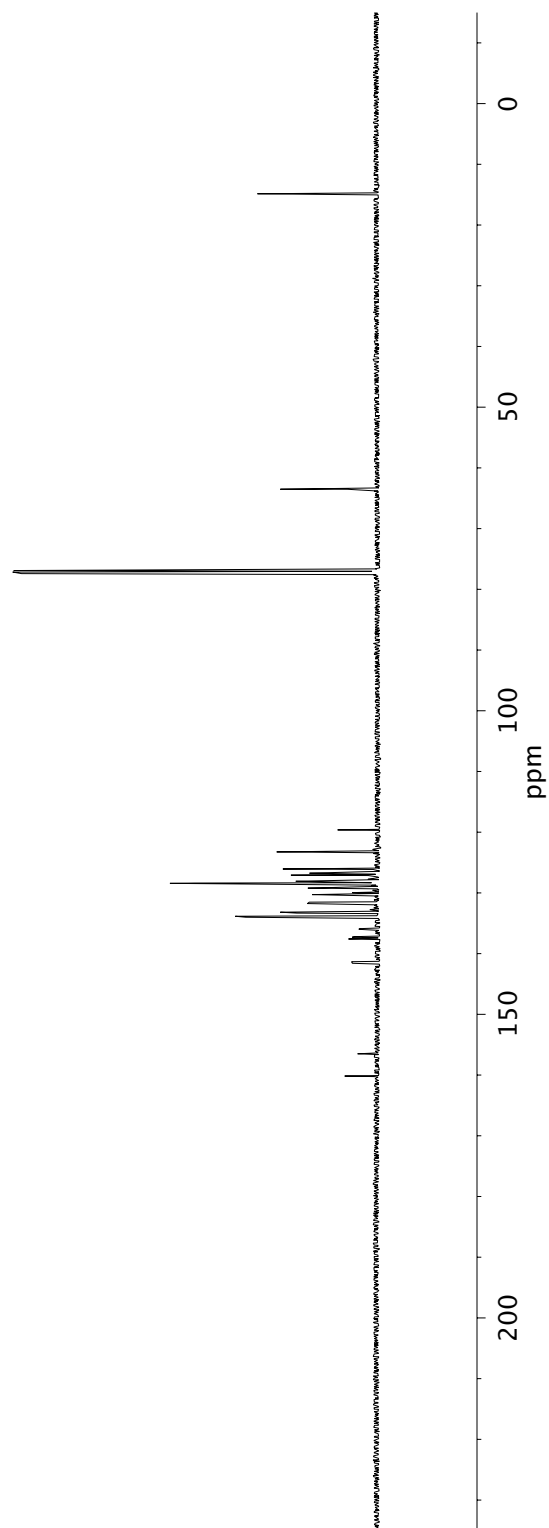
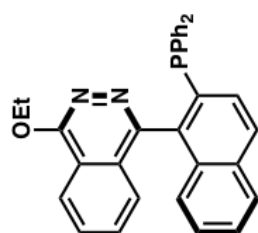


--14.26

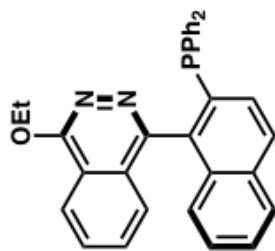


³¹P NMR (121 MHz, CDCl₃) of compound **4a**.

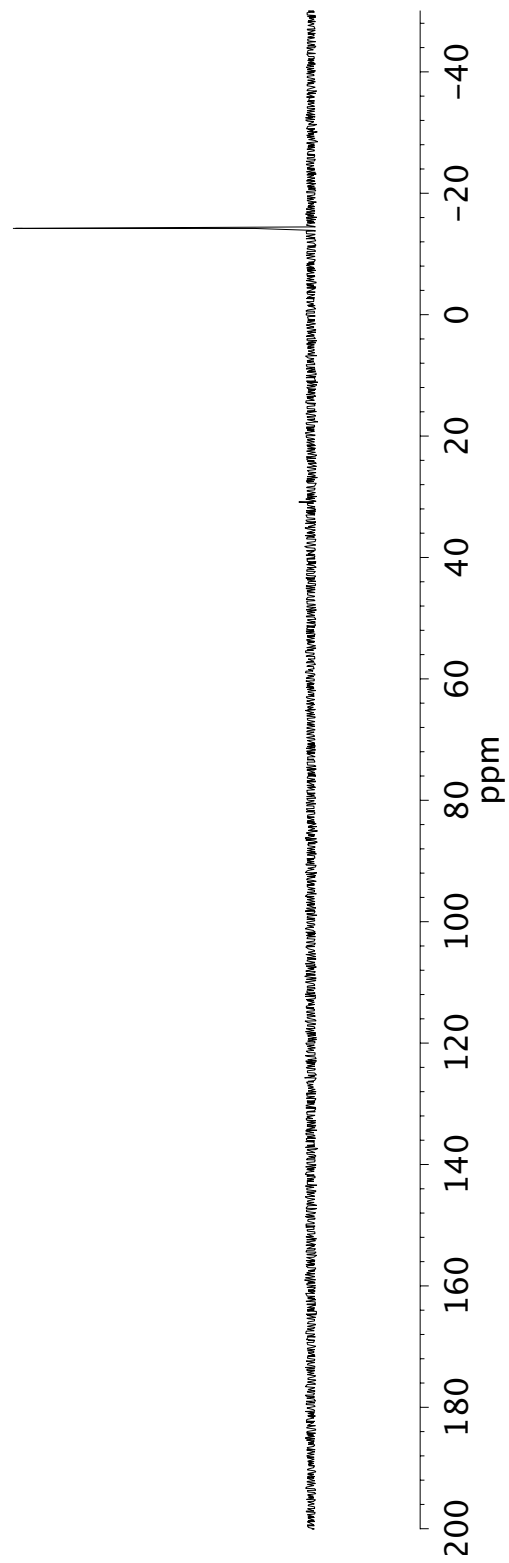
 ^1H NMR (500 MHz, CDCl_3) of compound **4b**.



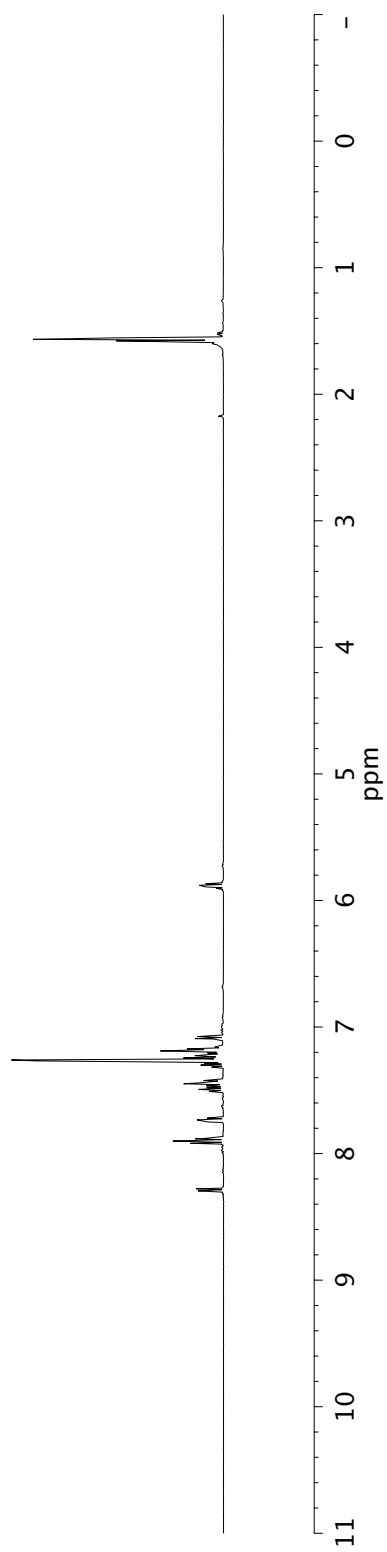
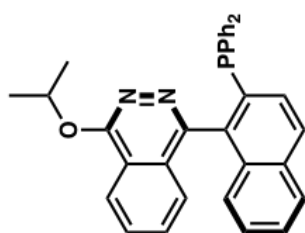
¹³C NMR (126 MHz, CDCl₃) of compound **4b**.



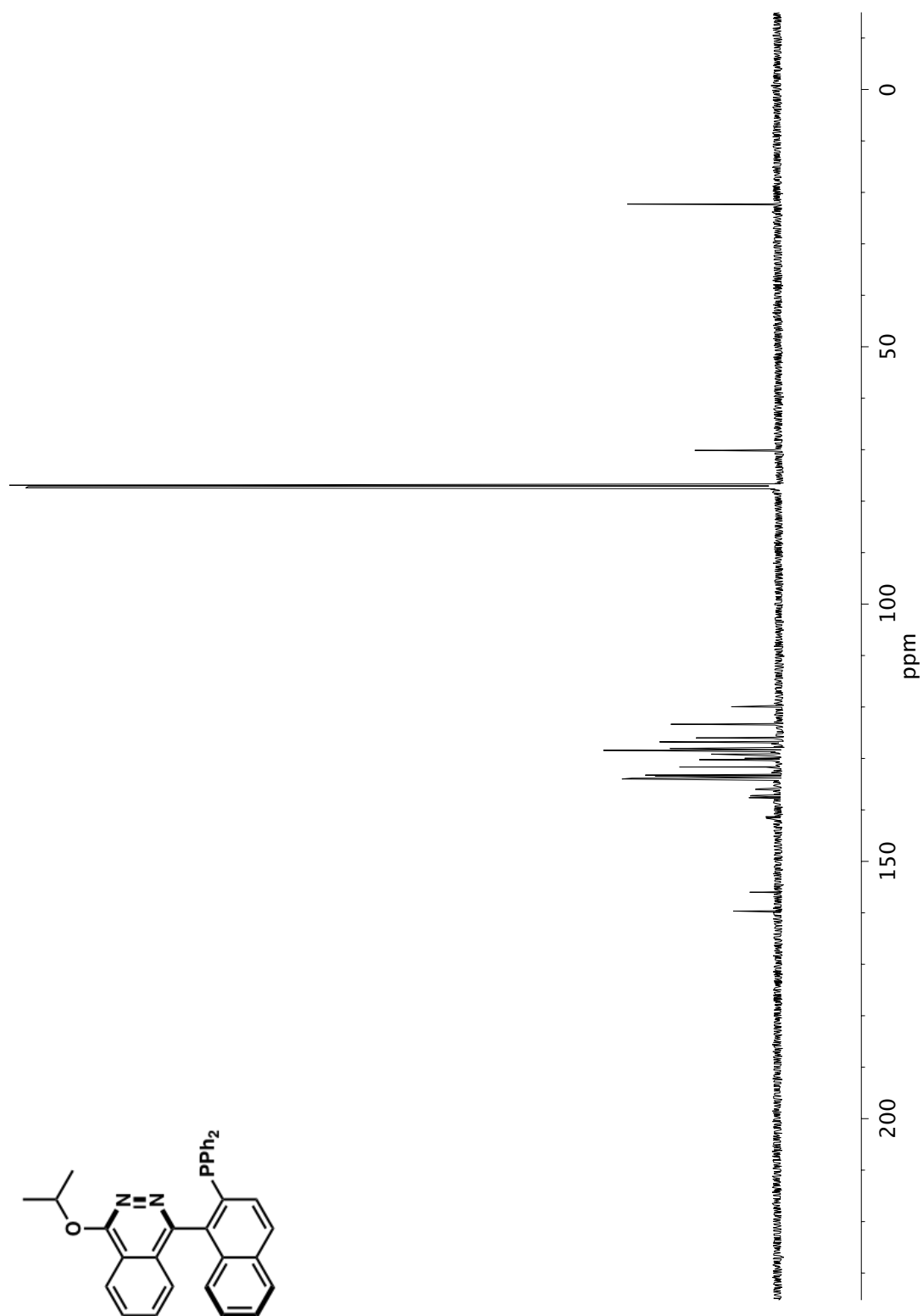
--14.21



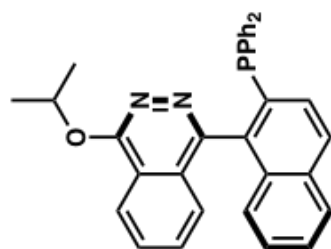
^{31}P NMR (121 MHz, CDCl_3) of compound **4b**.



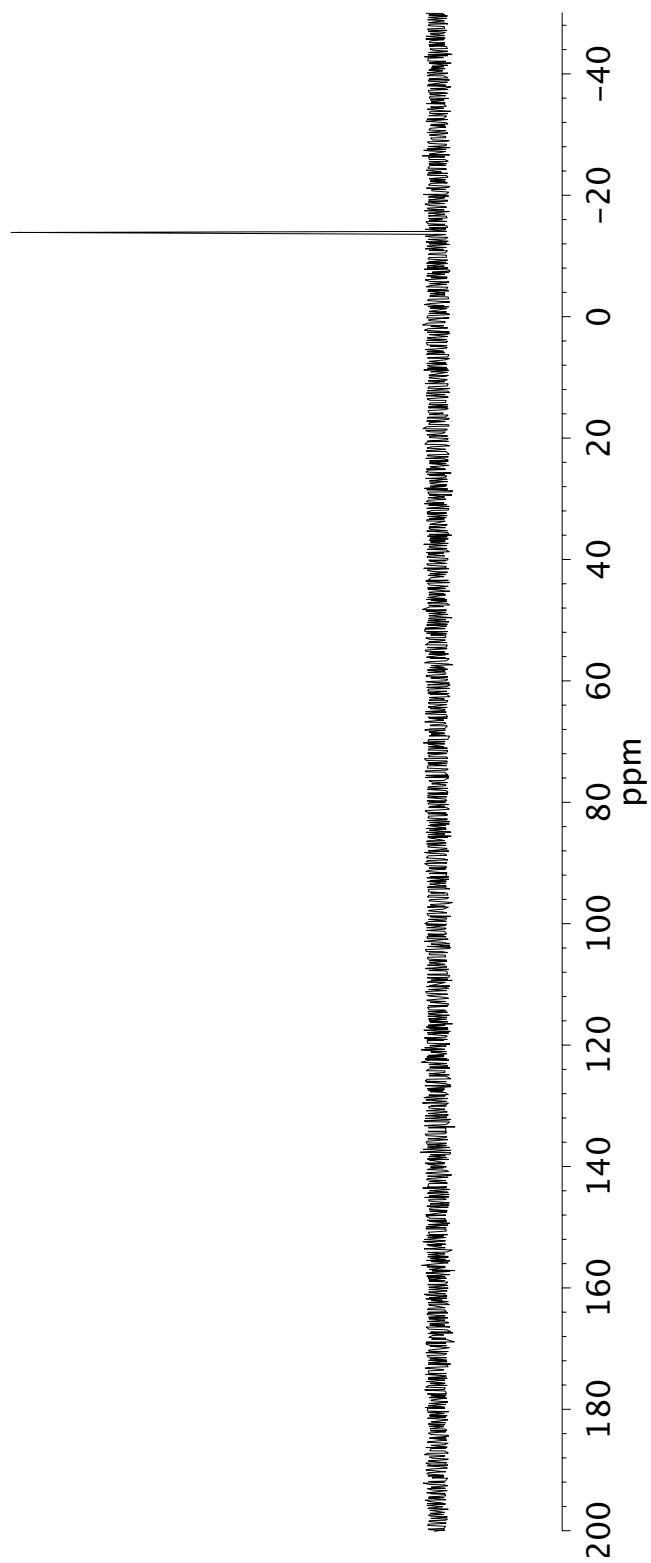
^1H NMR (500 MHz, CDCl_3) of compound **4c**.



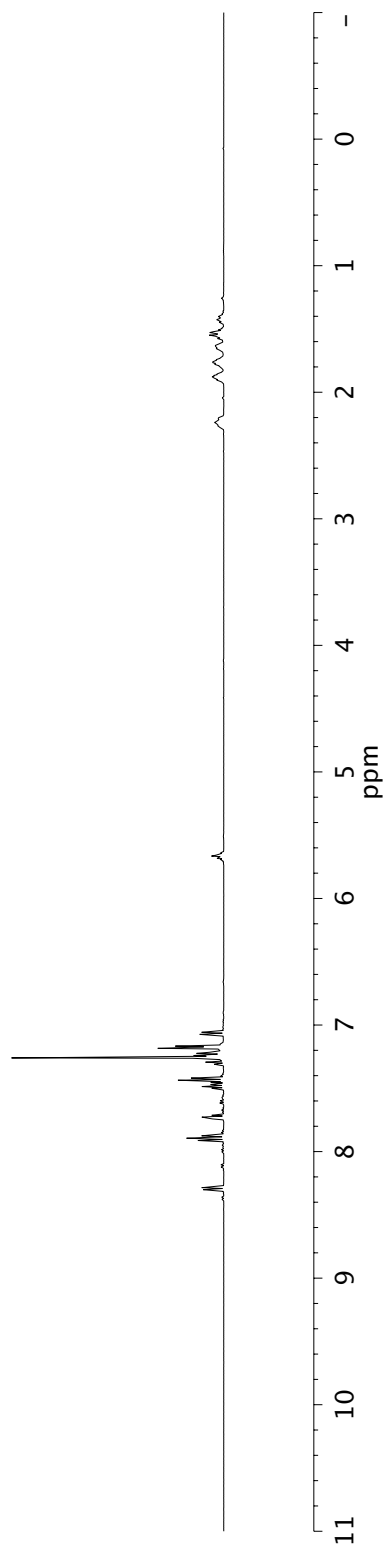
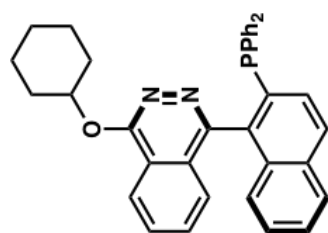
^{13}C NMR (126 MHz, CDCl_3) of compound **4c**.



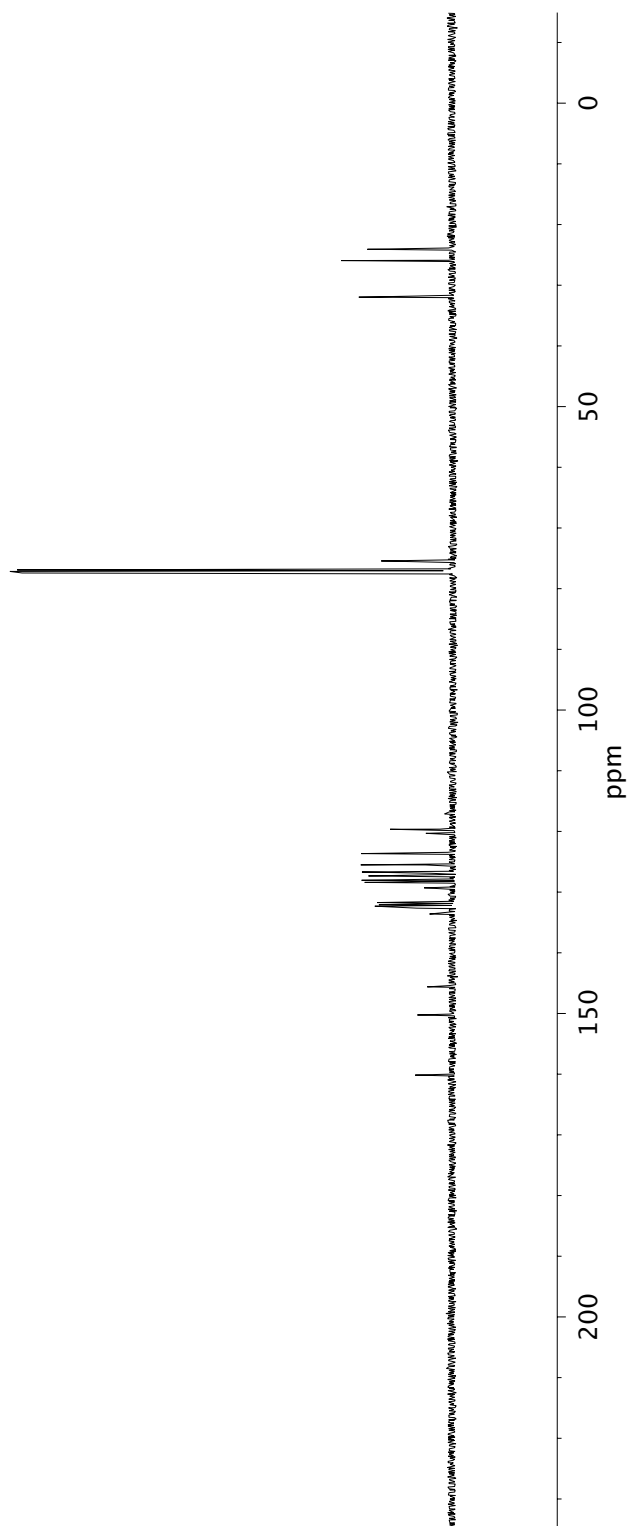
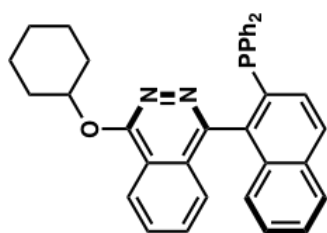
--13.88



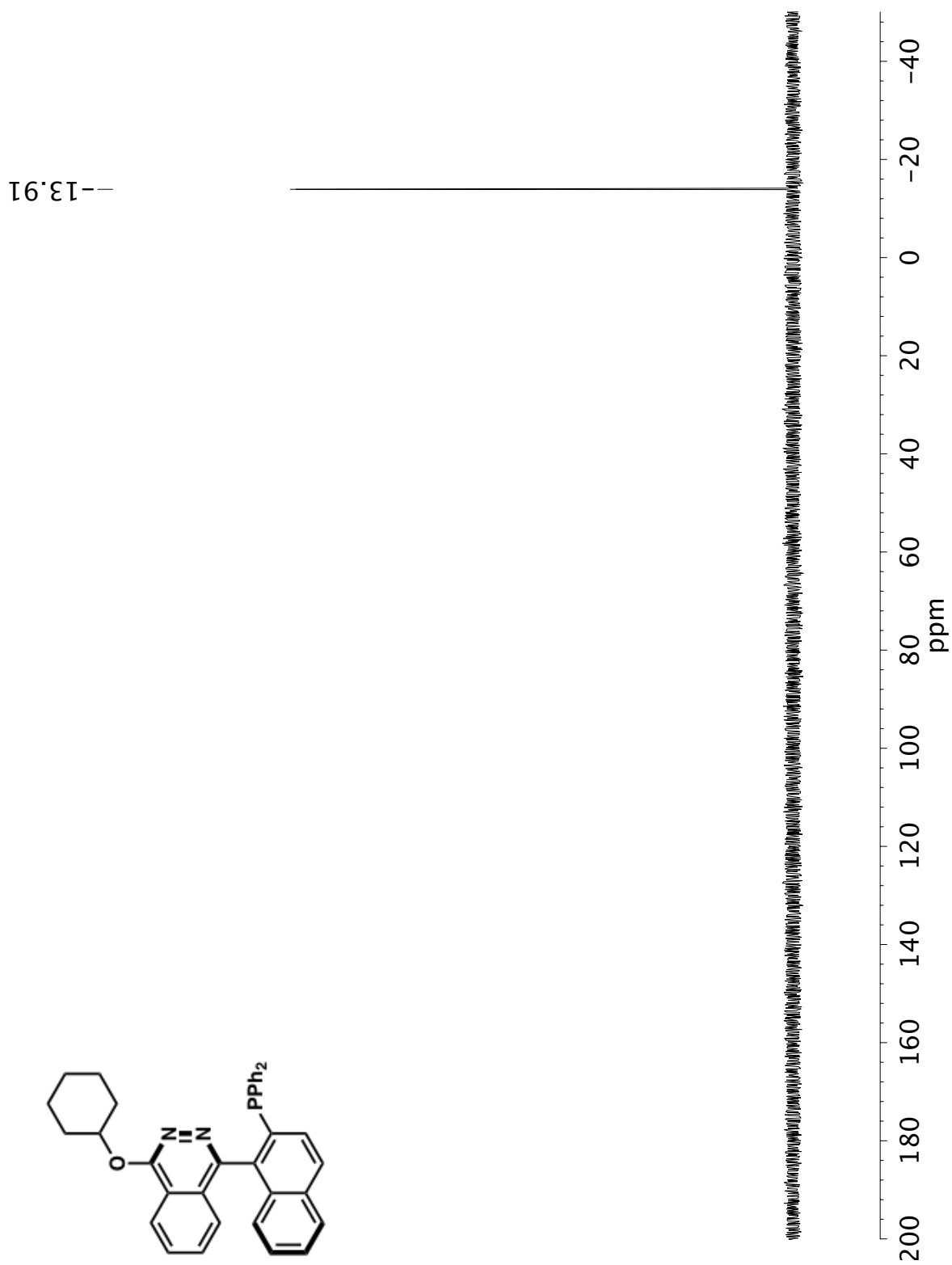
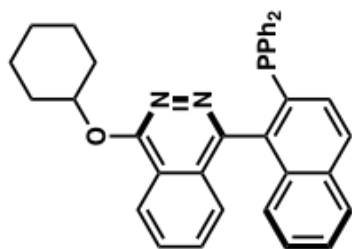
^{31}P NMR (121 MHz, CDCl_3) of compound **4c**.

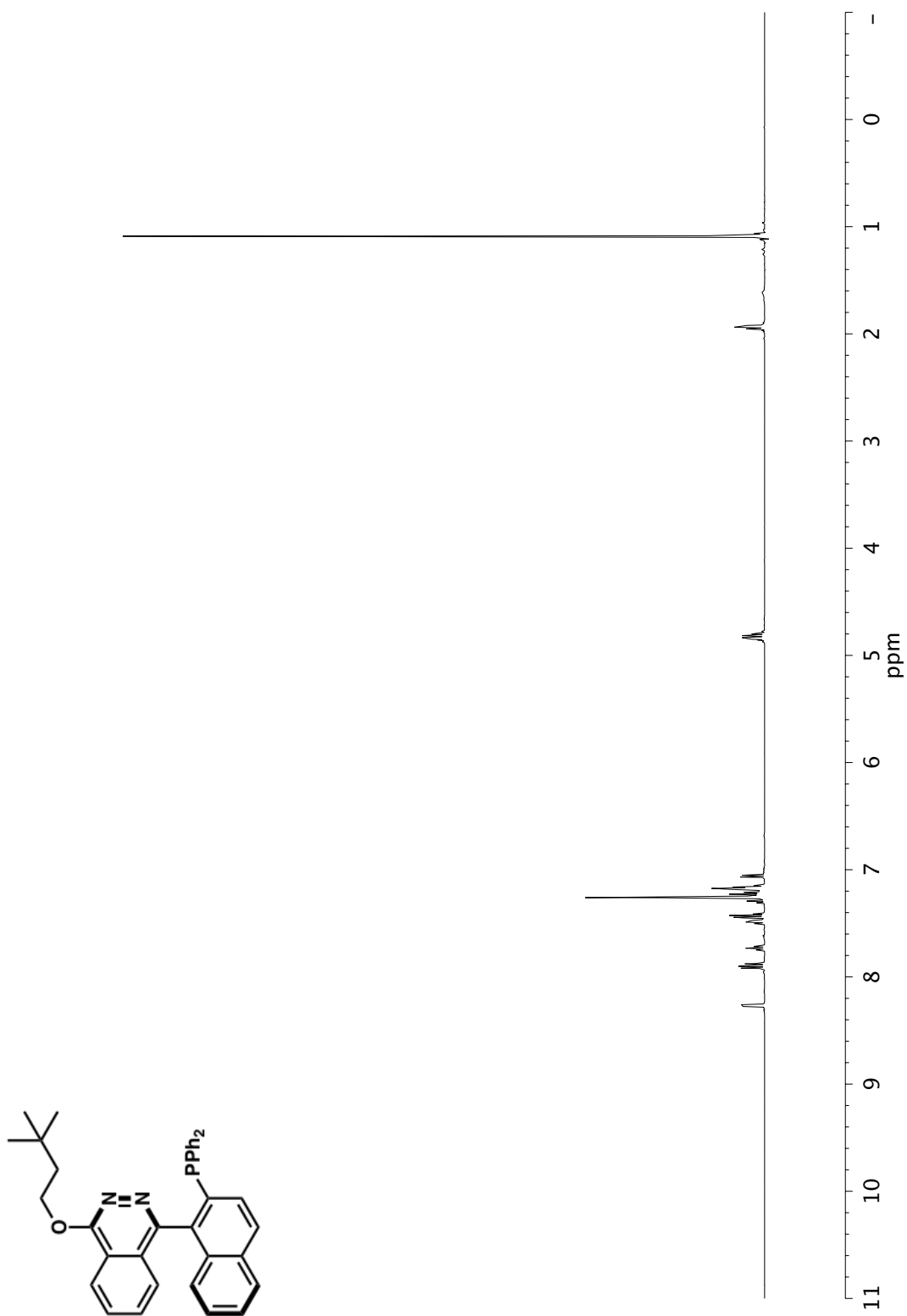


^1H NMR (500 MHz, CDCl_3) of compound **4d**.

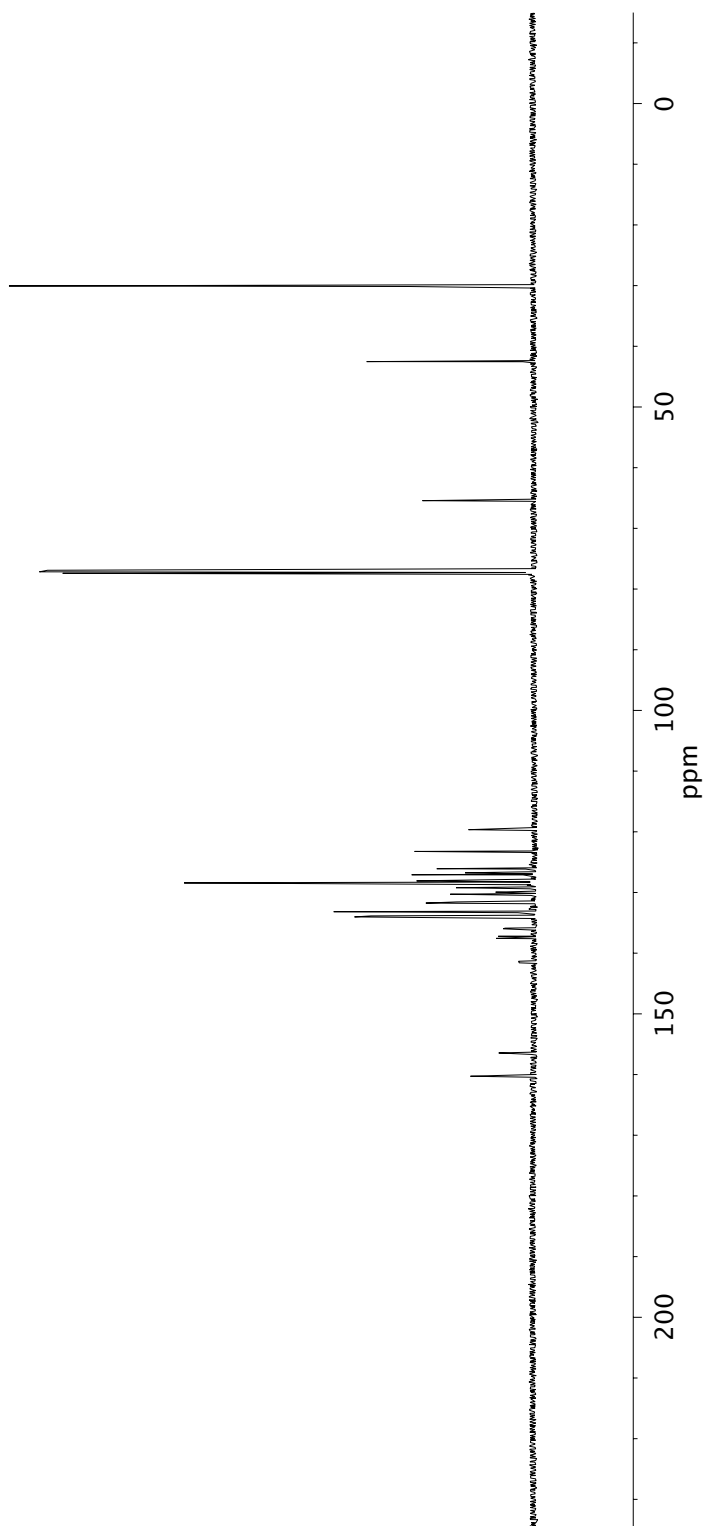
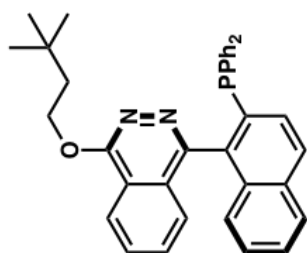


^{13}C NMR (126 MHz, CDCl_3) of compound **4d**.



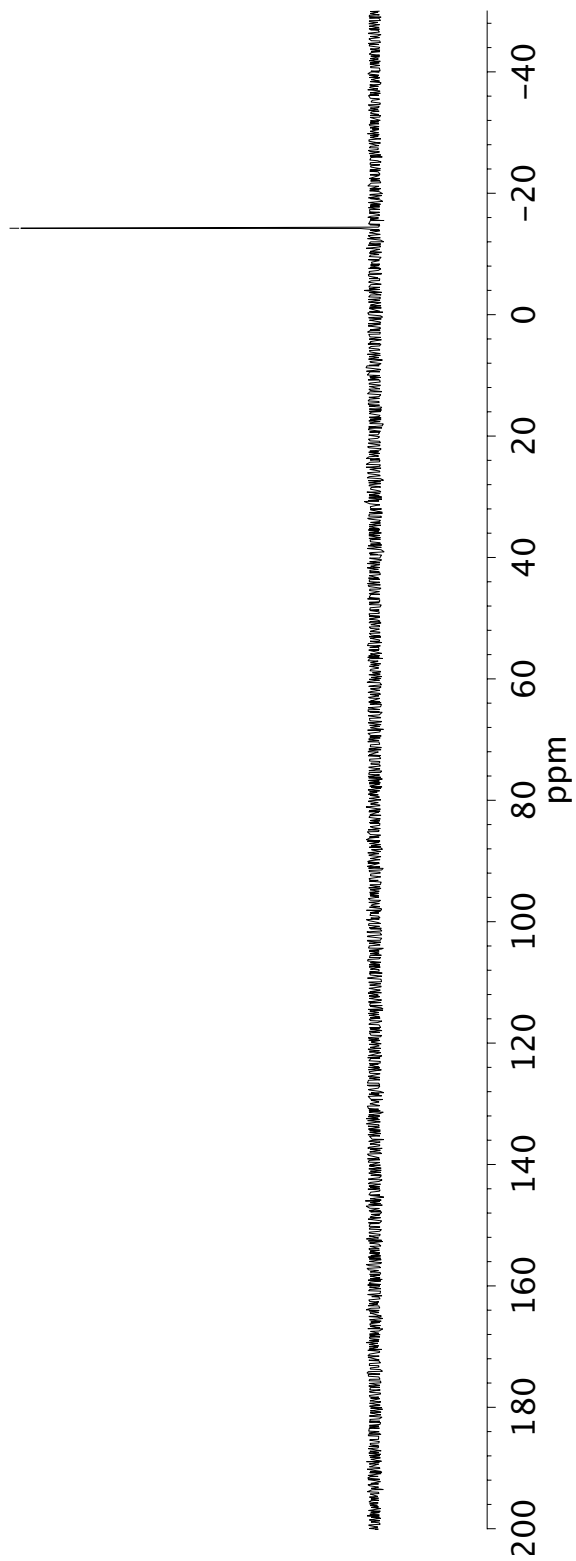
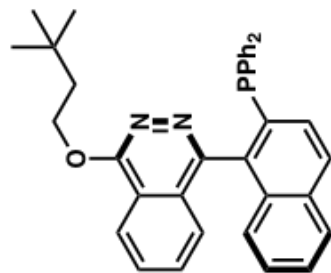


¹H NMR (500 MHz, CDCl₃) of compound **4e**.

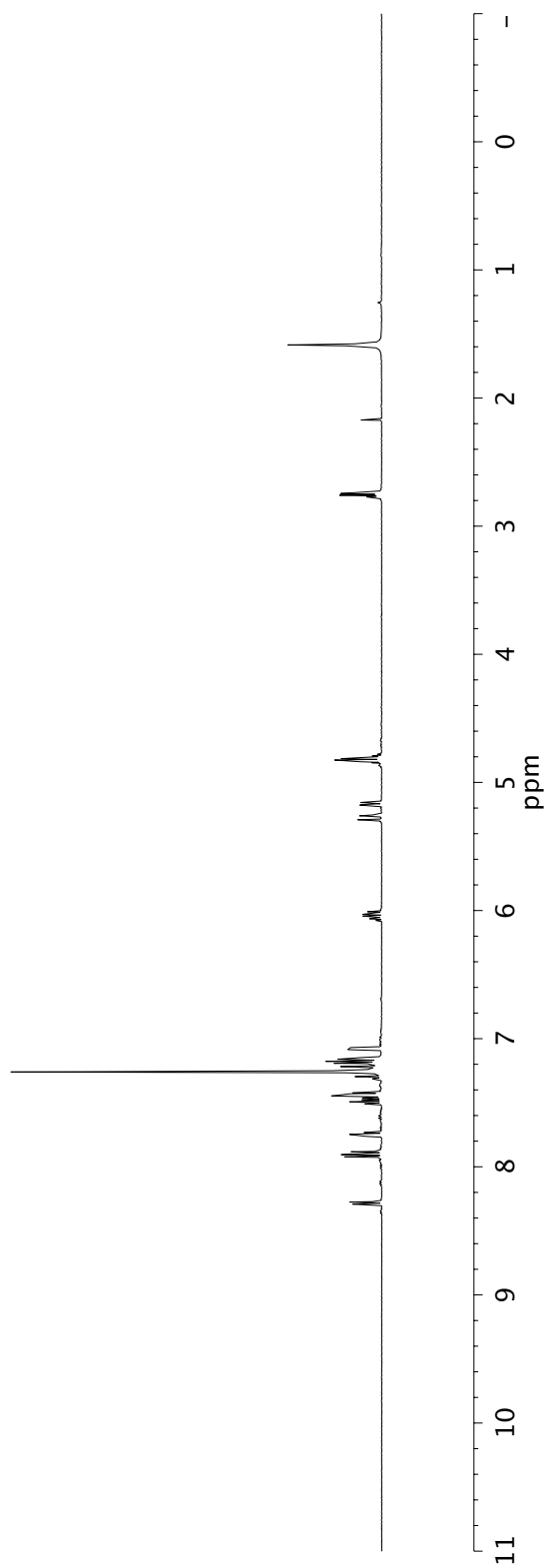
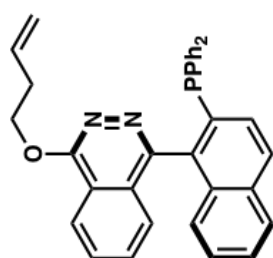


^{13}C NMR (126 MHz, CDCl_3) of compound **4e**.

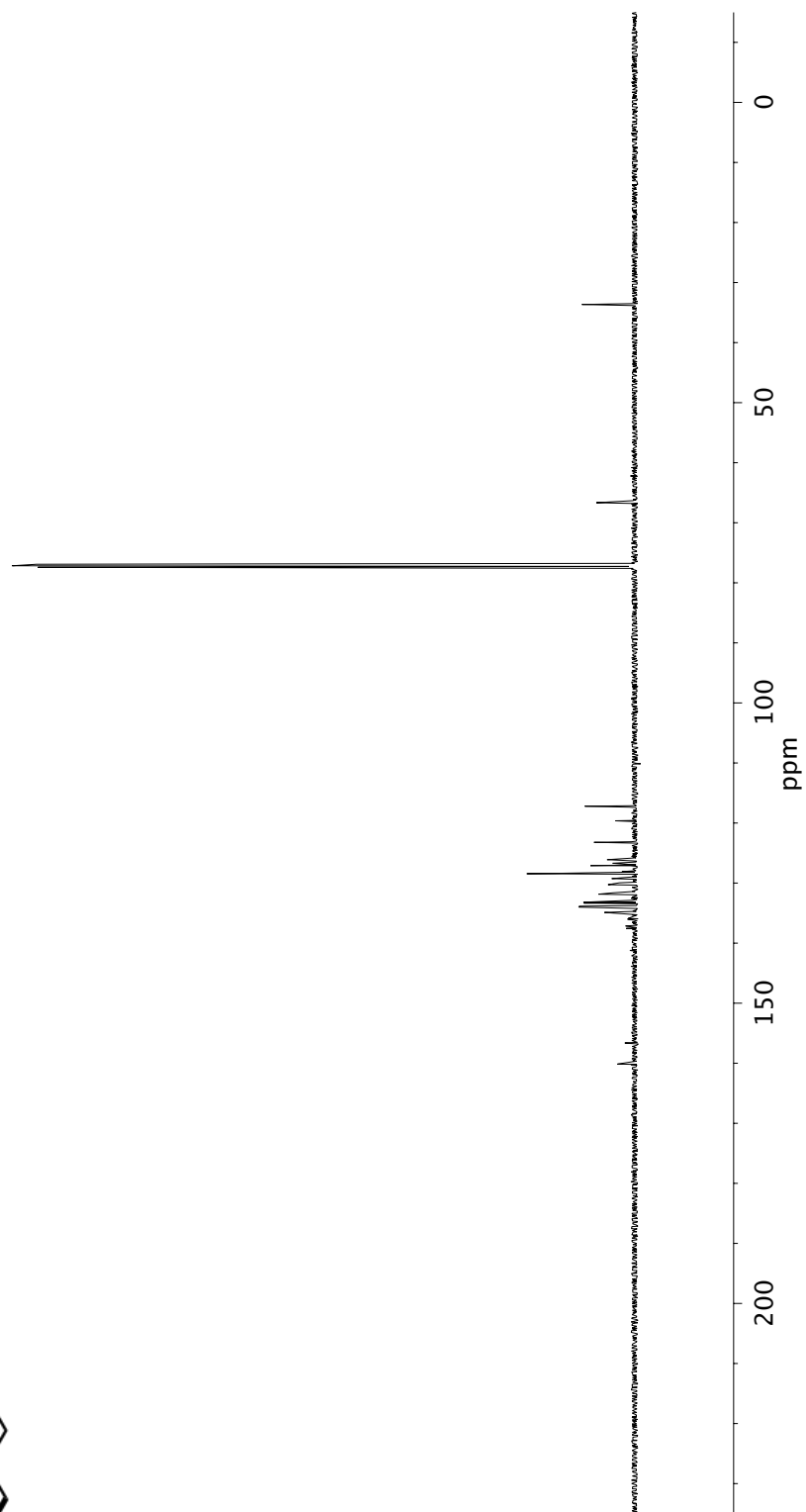
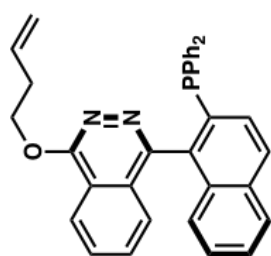
--14.24



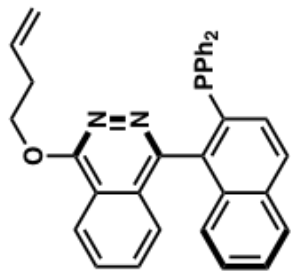
^{31}P NMR (121 MHz, CDCl_3) of compound **4e**.



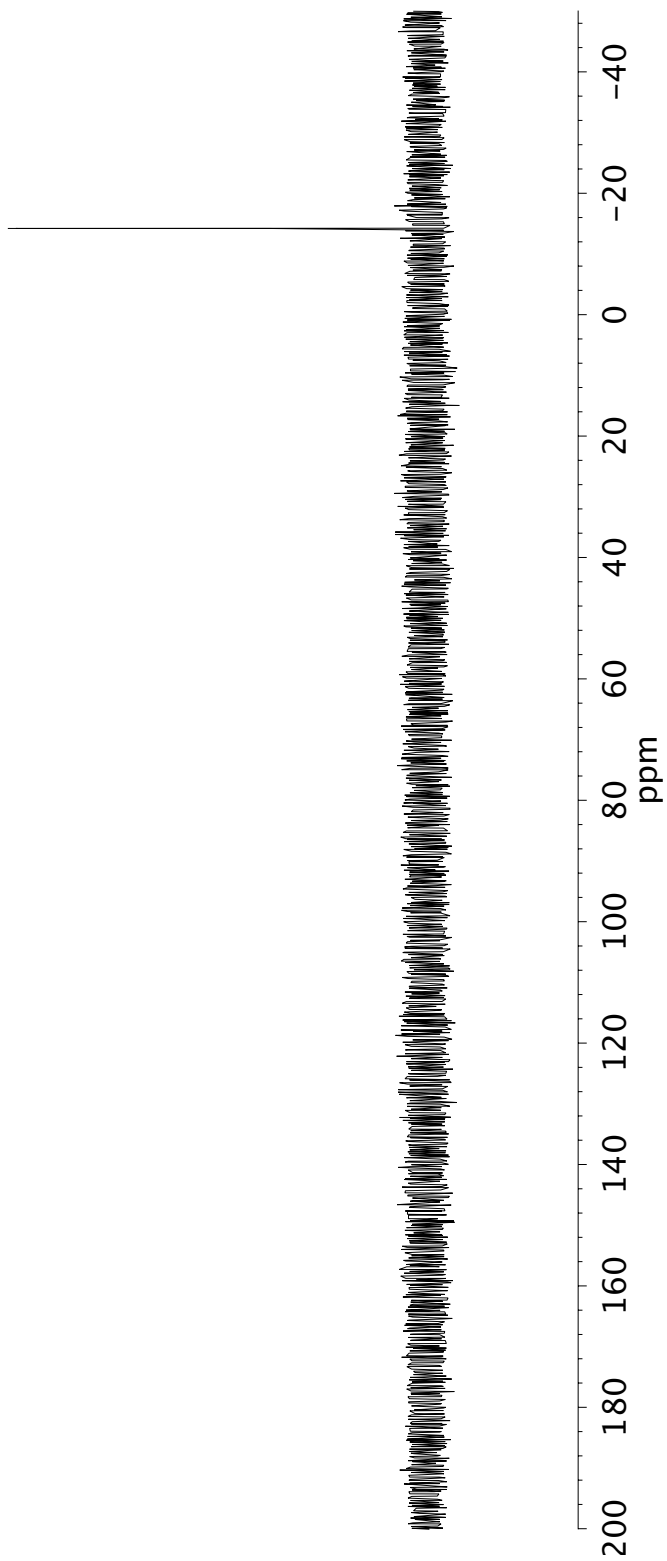
^1H NMR (500 MHz, CDCl_3) of compound **4f**.



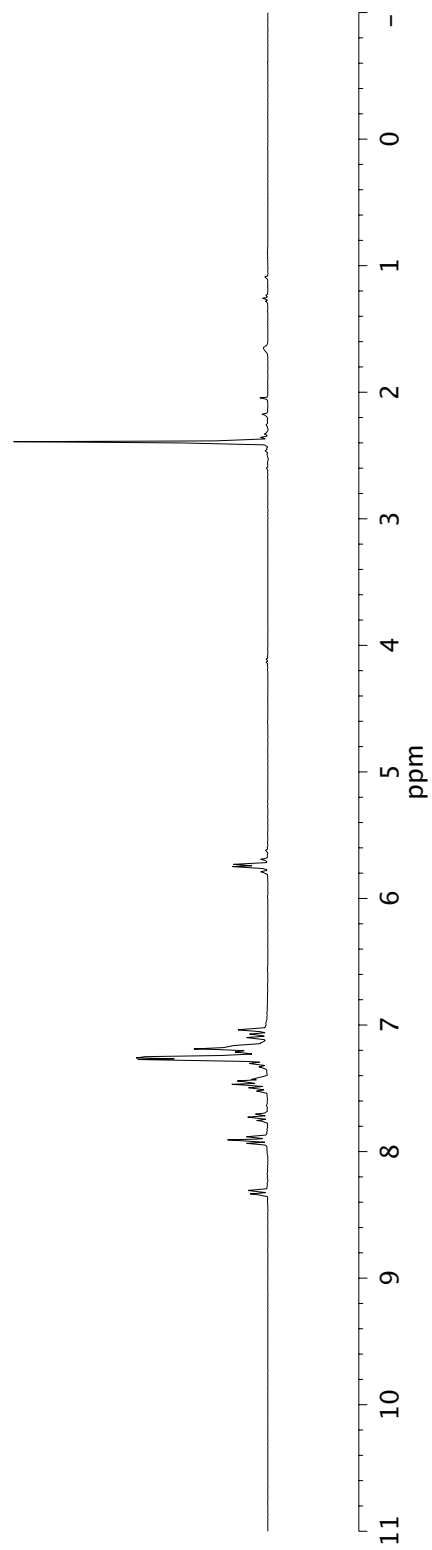
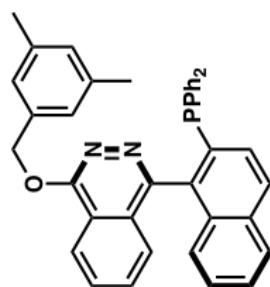
¹³C NMR (126 MHz, CDCl₃) of compound **4f**.



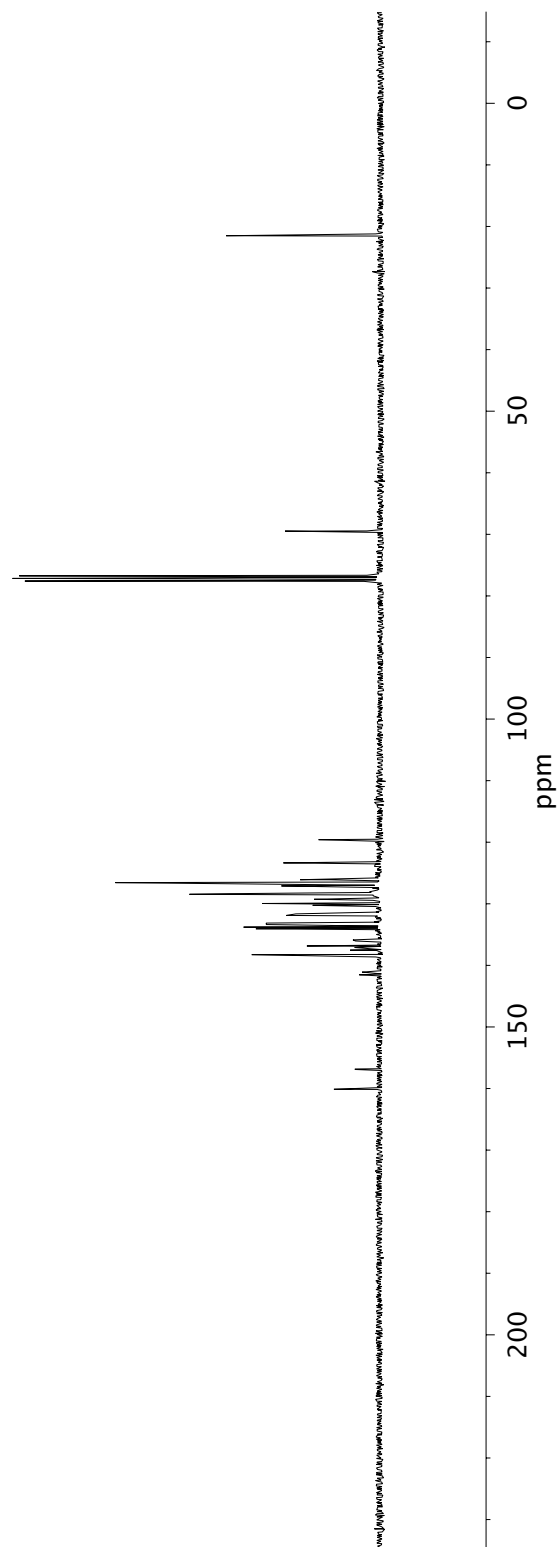
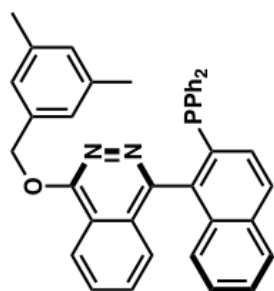
--14.20



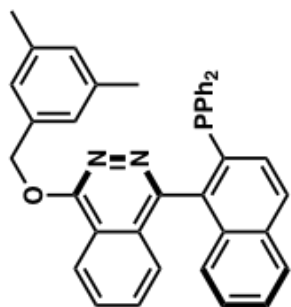
^{31}P NMR (121 MHz, CDCl_3) of compound 4f.



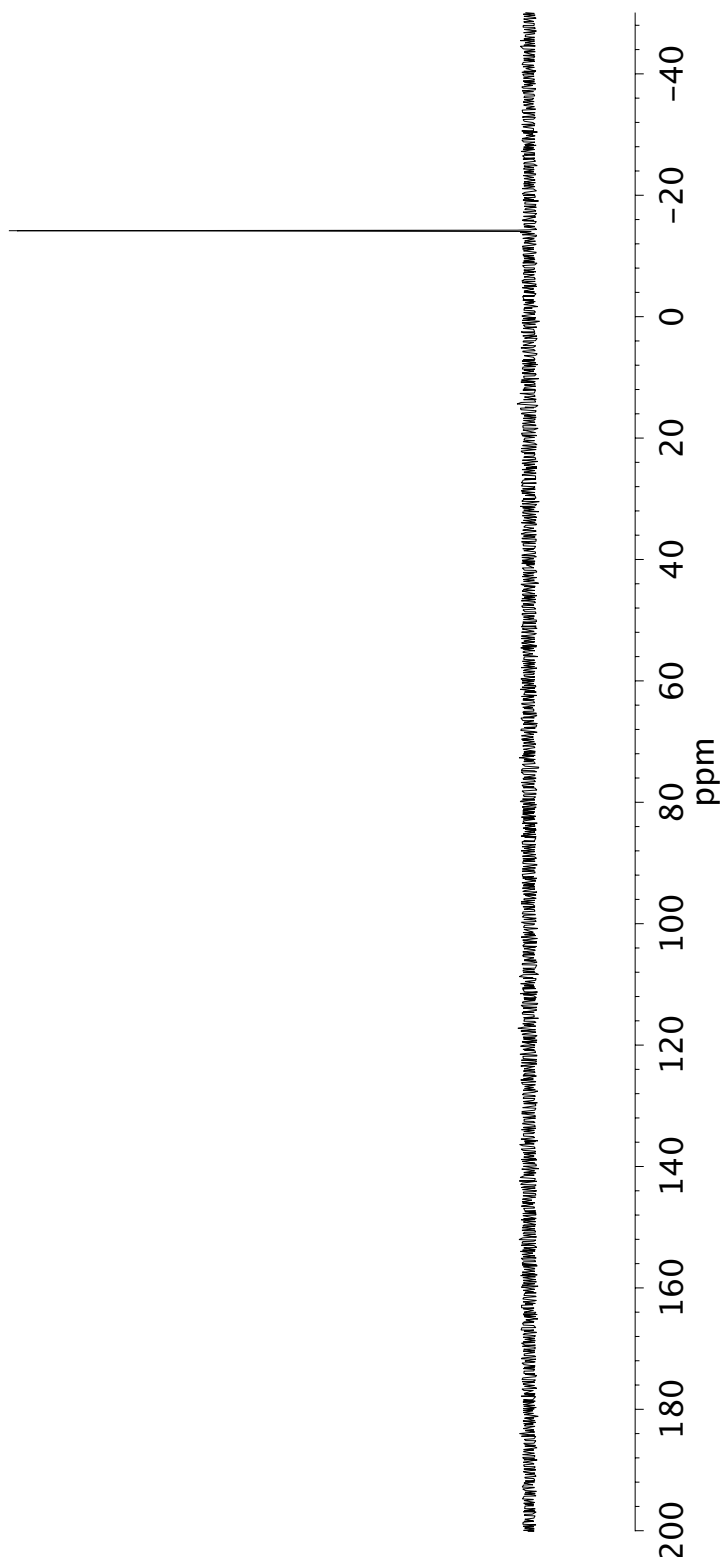
^1H NMR (500 MHz, CDCl_3) of compound **4g**.



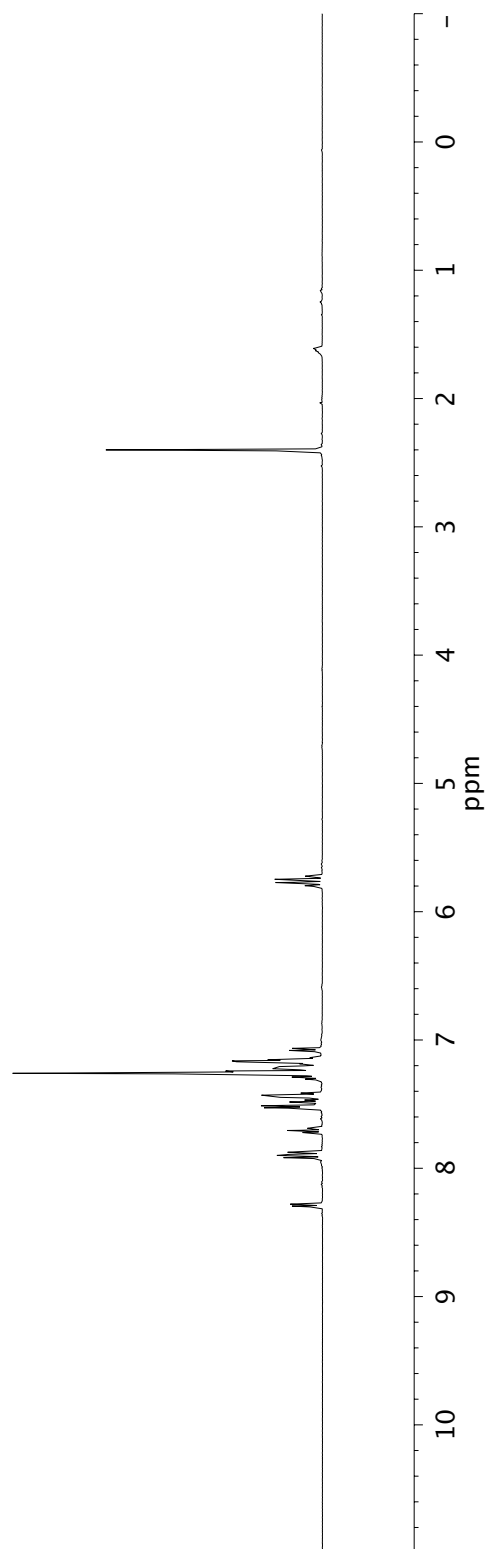
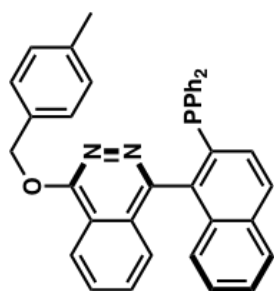
^{13}C NMR (126 MHz, CDCl_3) of compound 4g.



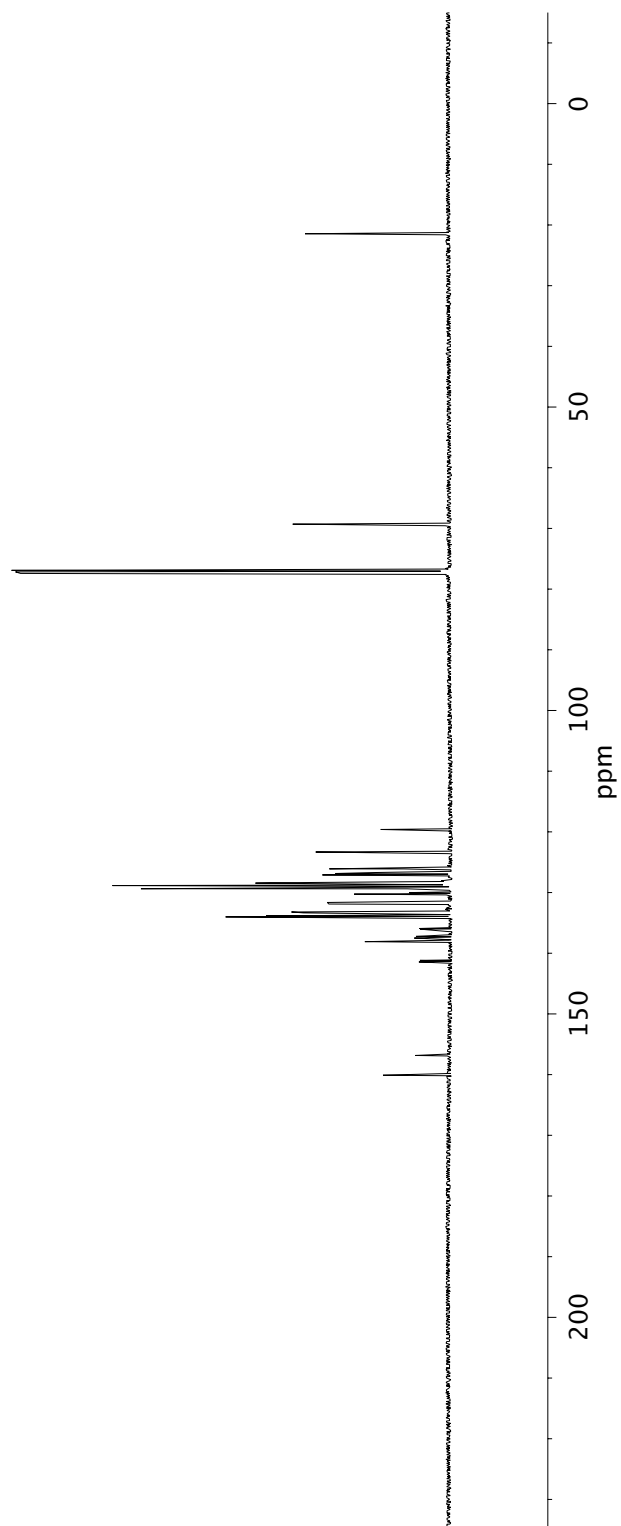
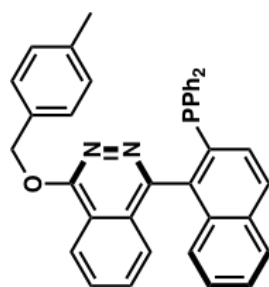
--14.16



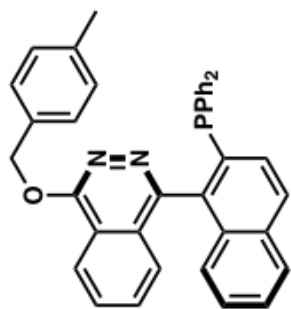
³¹P NMR (121 MHz, CDCl₃) of compound **4g**.



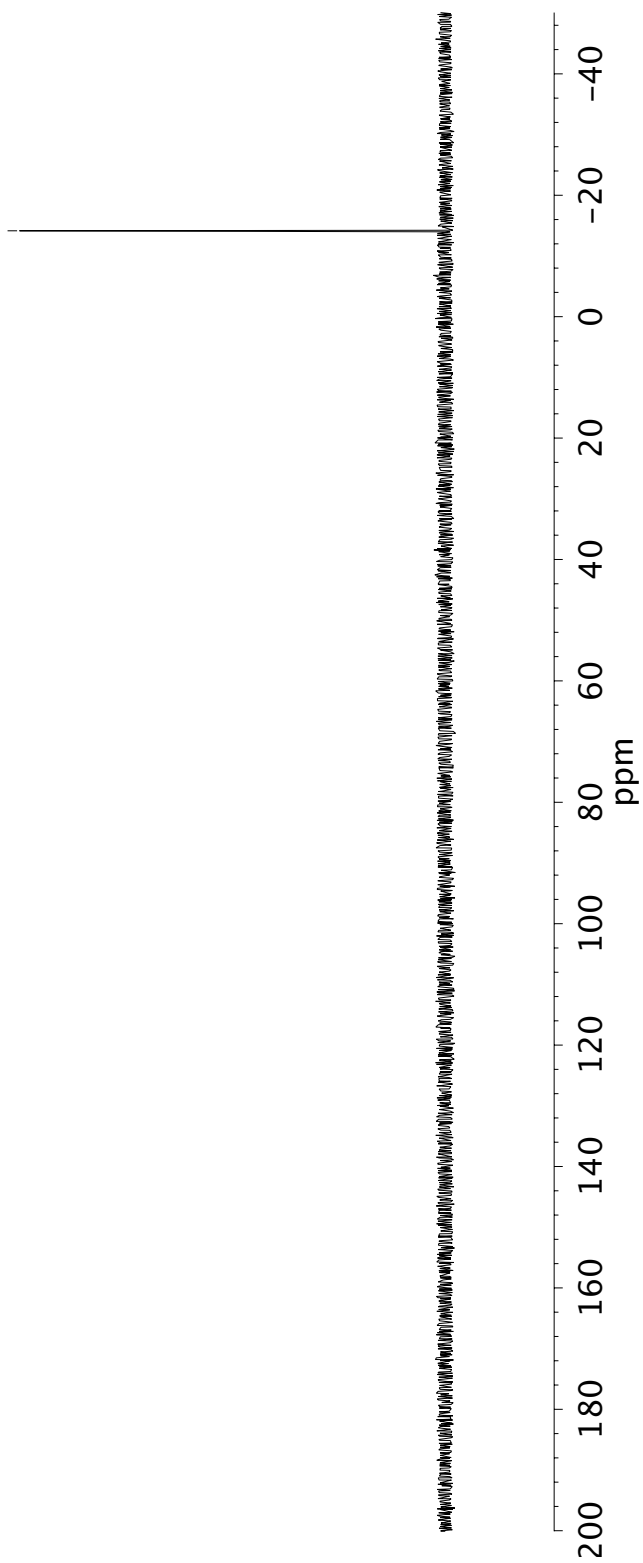
^1H NMR (500 MHz, CDCl_3) of compound **4h**.



^{13}C NMR (126 MHz, CDCl_3) of compound **4h**.



--14.15

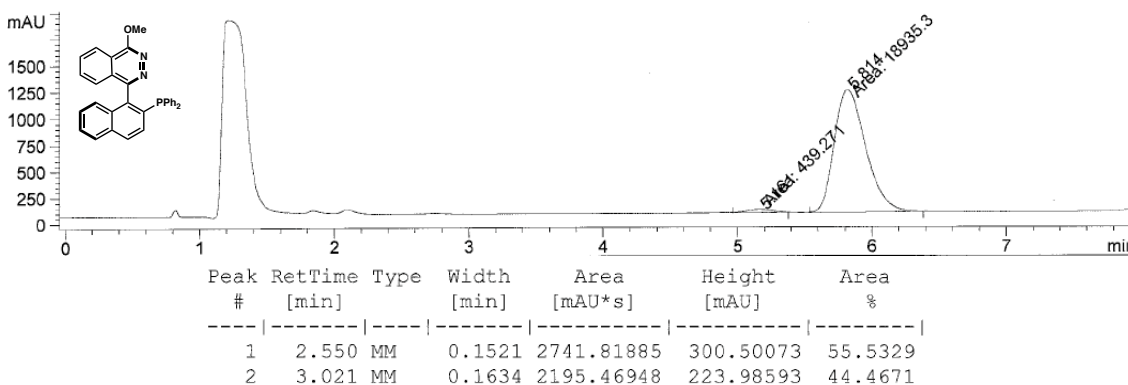
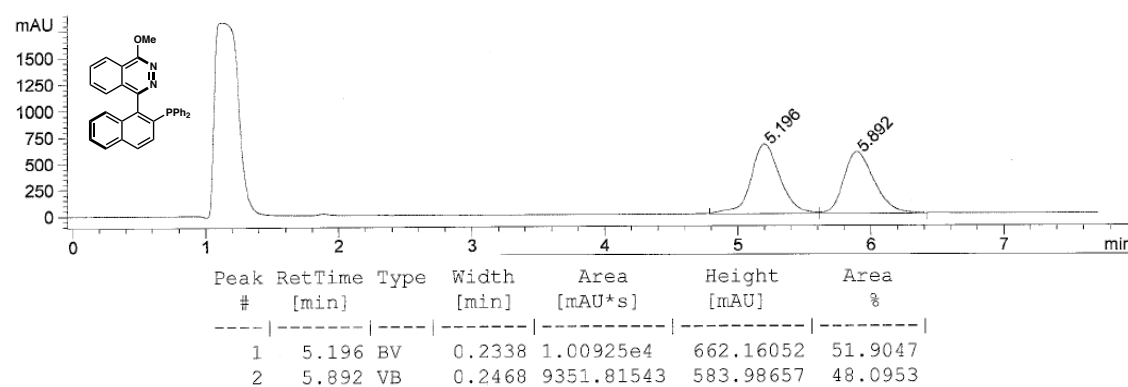


^{31}P NMR (121 MHz, CDCl_3) of compound **4h**.

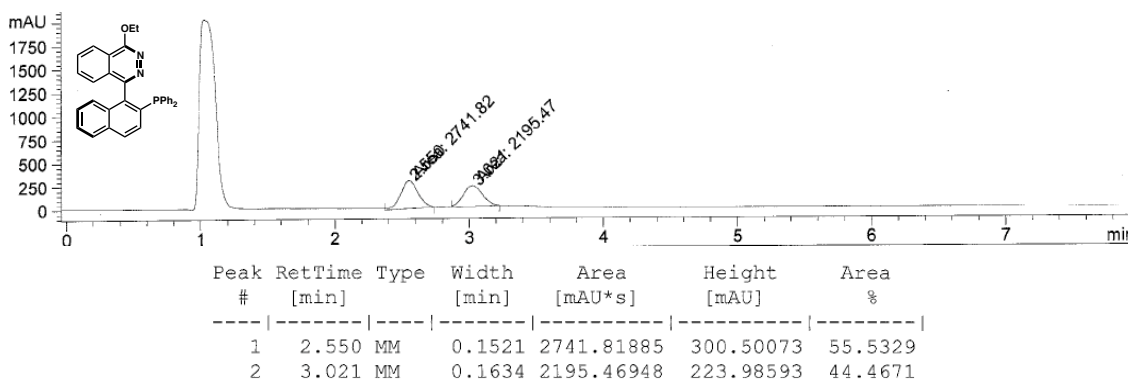
Copies of SFC/HPLC spectra

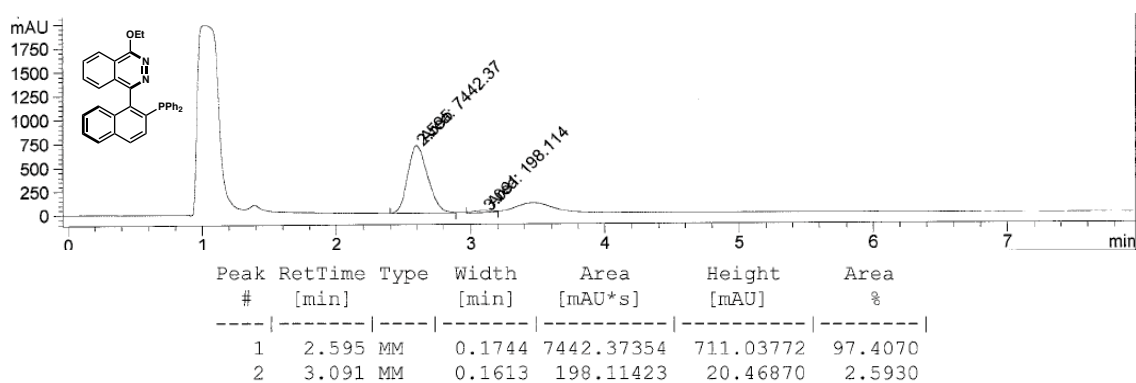
(R)-1-(2-(diphenylphosphaneyl)naphthalen-1-yl)-4-methoxyphthalazine (**4a**)

SFC conditions: 35% IPA, 2.5mL/min, Chiralcel OD-H column, t_R (min): major = 5.81, minor = 5.16.

*(R)*-1-(2-(diphenylphosphaneyl)naphthalen-1-yl)-4-ethoxyphthalazine (**4b**)

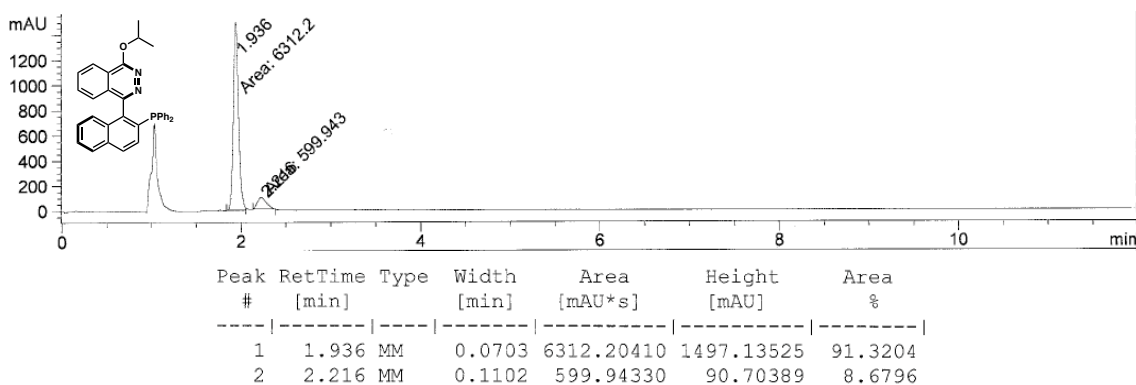
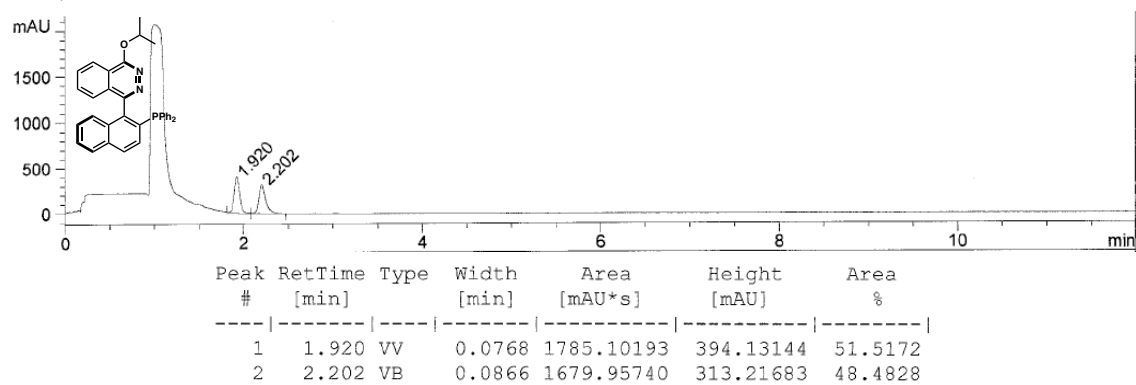
SFC conditions: 30% MeOH, 2.5mL/min, Chiralpak AD-H column, t_R (min): major = 2.60, minor = 3.09.





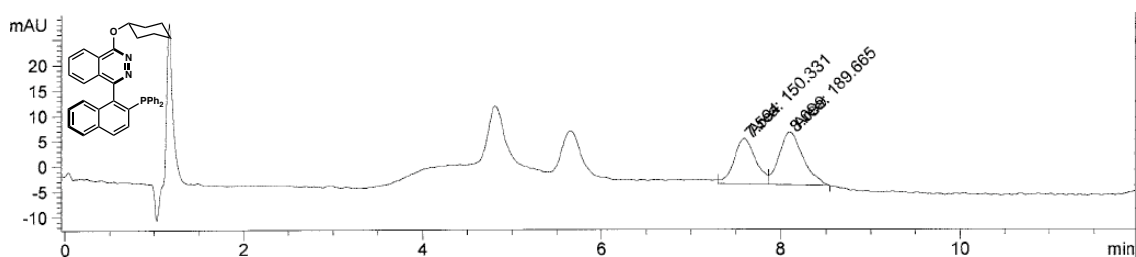
(R)-1-(2-(diphenylphosphaneyl)naphthalen-1-yl)-4-isopropoxyphthalazine (4c)

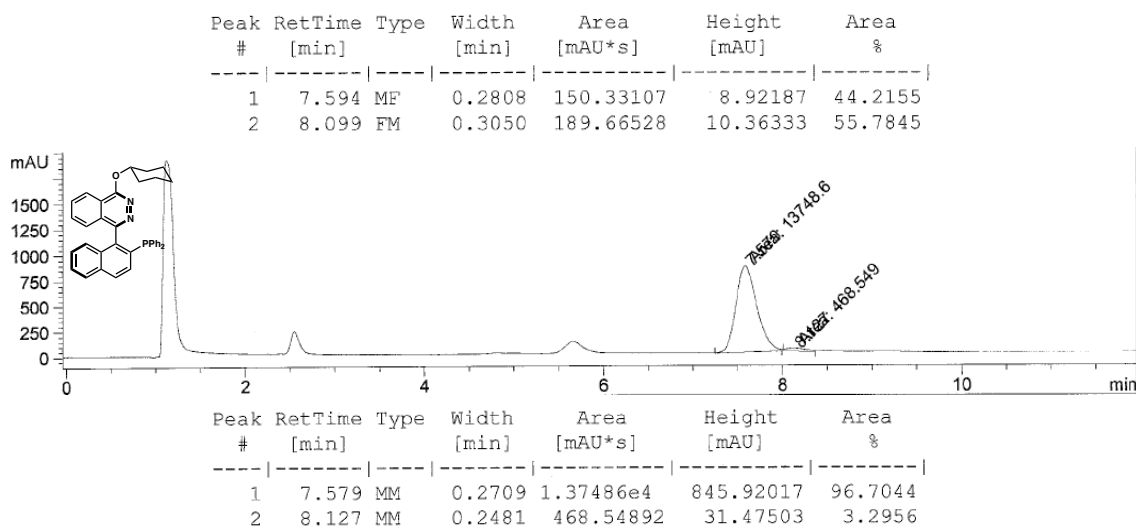
SFC conditions: 40% MeOH, 2.5mL/min, Chiralpak AD-H column, t_R (min): major = 1.94, minor = 2.22.



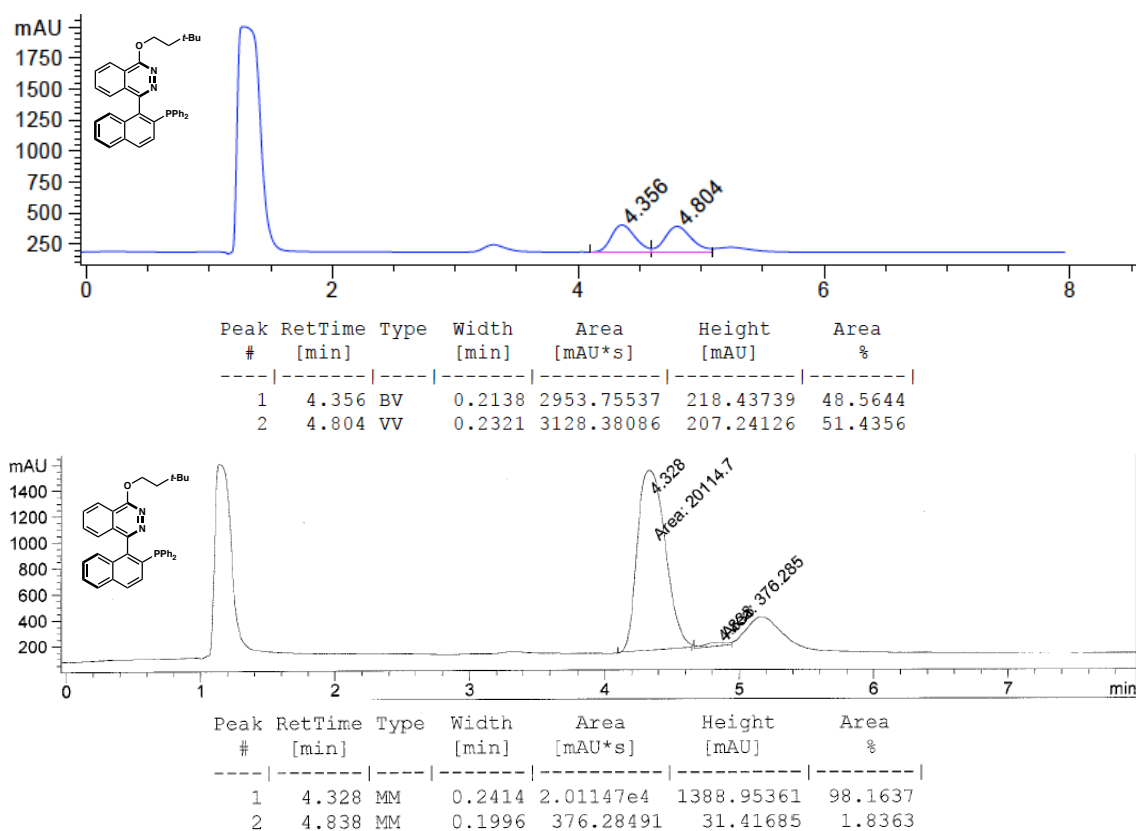
(R)-1-(cyclohexyloxy)-4-(2-(diphenylphosphaneyl)naphthalen-1-yl)phthalazine (4d)

SFC conditions: 25% MeOH, 2.5mL/min, Chiralcel OD-H column, t_R (min): major = 7.58, minor = 8.13.



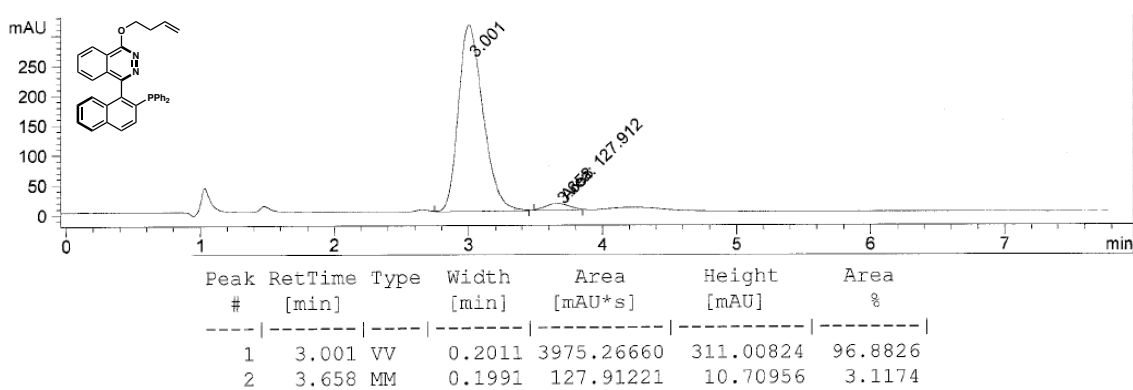
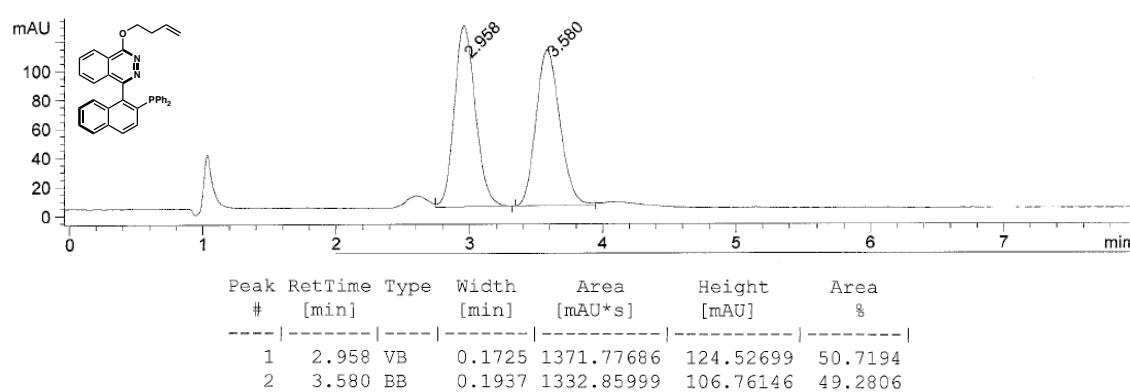


(*R*)-1-(3,3-dimethylbutoxy)-4-(2-(diphenylphosphaneyl)naphthalen-1-yl)phthalazine (**4e**)
 SFC conditions: 35% IPA, 2.5mL/min, Chiralcel OD-H column, t_R (min): major = 4.33, minor = 4.84.



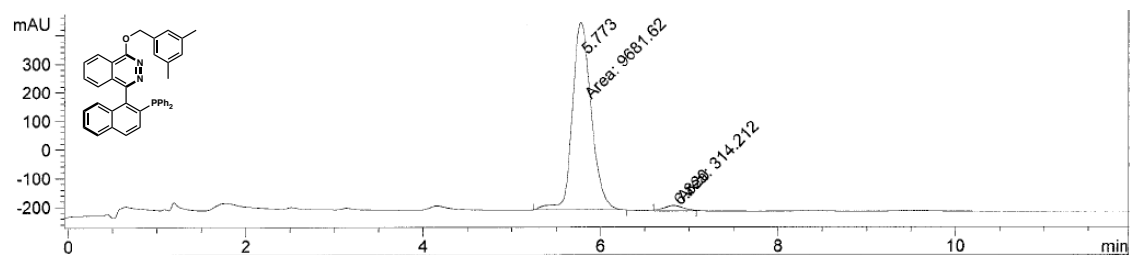
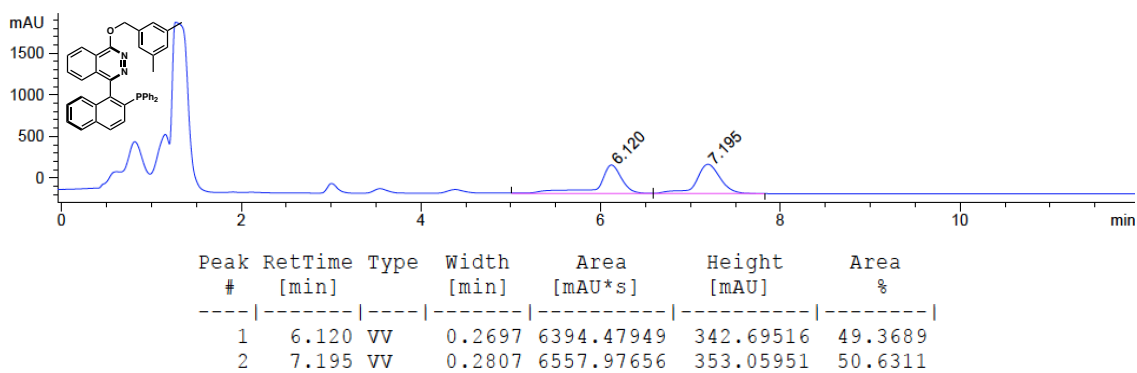
(*R*)-1-(but-3-en-1-yloxy)-4-(2-(diphenylphosphaneyl)naphthalen-1-yl)phthalazine (**4f**)
 SFC conditions: 35% MeOH, 2.5mL/min, Chiralpak AD-H column, t_R (min): major =

3.00, minor = 3.66.



(*R*)-1-((3,5-dimethylbenzyl)oxy)-4-(2-(diphenylphosphaneyl)naphthalen-1-yl)phthalazine (**4g**)

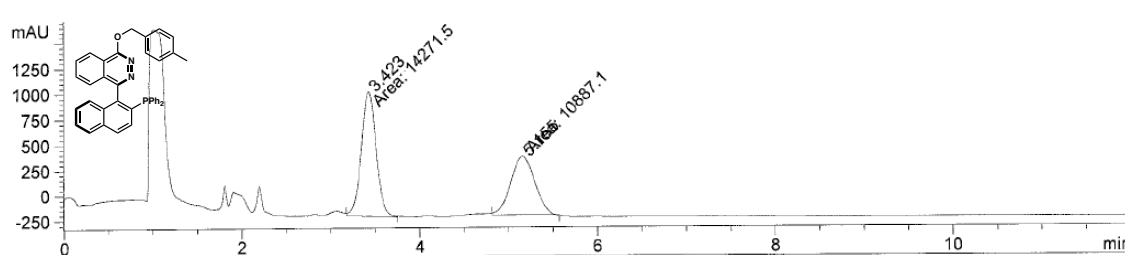
SFC conditions: 40% MeOH, 2.5mL/min, Chiralcel OD-H column, t_R (min): major = 5.77, minor = 6.82.



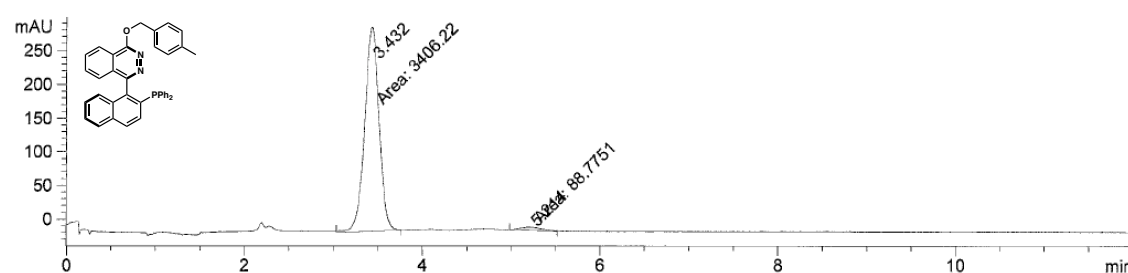
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	5.773	MM	0.2476	9681.61914	651.68408	96.8566
2	6.820	MM	0.2737	314.21243	19.13087	3.1434

(R)-1-(2-(diphenylphosphaneyl)naphthalen-1-yl)-4-((4-methylbenzyl)oxy)phthalazine (4h)

SFC conditions: 40% MeOH, 2.5mL/min, Chiralpak AD-H column, t_R (min): major = 3.43, minor = 5.21.



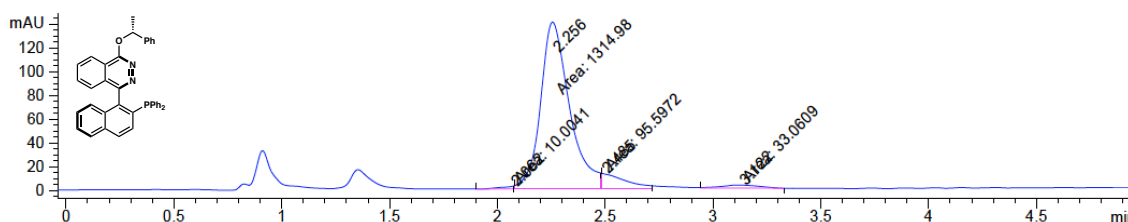
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	3.423	MM	0.1945	1.42715e4	1223.03918	56.7261
2	5.155	MM	0.3141	1.08871e4	577.68909	43.2739



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	3.432	MM	0.1881	3406.21655	301.85727	97.4599
2	5.214	MM	0.2921	88.77509	5.06575	2.5401

1-((R)-2-(diphenylphosphaneyl)naphthalen-1-yl)-4-((R)-1-phenylethoxy)phthalazine (4i)

SFC conditions: 40% IPA, 2.5 mL/min, Chiralpak OD-H column, t_R (min): major = 2.26, minor = 3.12.



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	2.062	MM	0.0871	10.00407	1.91379	0.6882
2	2.256	MM	0.1553	1314.98193	141.13321	90.4611
3	2.485	MM	0.1229	95.59724	12.96553	6.5764
4	3.122	MM	0.2031	33.06094	2.71275	2.2743

1-(2-(diphenylphosphaneyl)naphthalen-1-yl)-4-((*R*)-1-phenylethoxy)phthalazine (**2a**)

HPLC conditions: 5% IPA/hexane, 1.5 mL/min, Chiralpak OD-H column, tR (min): major = 2.91, minor = 3.47. (prior to crystallization)

